

# The Role of Inflammatory Mediators in the Pathogenesis of Alzheimer's Disease

Gholamreza Azizi,<sup>1,2</sup> Shadi S. Navabi,<sup>3</sup> Ahmed Al-Shukaili,<sup>4</sup> Mir H. Seyedzadeh,<sup>3</sup> Reza Yazdani,<sup>2,5</sup> \*Abbas Mirshafiey<sup>3</sup>

## دور وسائط الالتهاب في الآلية المرضية لداء الزهايمر

غلامريزا عزيزي، شادي سادات نافابي، أحمد الشكيلي، مير هادي سيدزاده، رضا يزدي، عباس ميرشافي

**ABSTRACT:** Alzheimer's disease (AD), a neurodegenerative disorder associated with advanced age, is the most common cause of dementia globally. AD is characterised by cognitive dysfunction, deposition of amyloid plaques, neurofibrillary tangles and neuro-inflammation. Inflammation of the brain is a key pathological hallmark of AD. Thus, clinical and immunopathological evidence of AD could be potentially supported by inflammatory mediators, including cytokines, chemokines, the complement system, acute phase proteins and oxidative mediators. In particular, oxidative mediators may actively contribute to the progression of AD and on-going inflammation in the brain. This review provides an overview of the functions and activities of inflammatory mediators in AD. An improved understanding of inflammatory processes and their role in AD is needed to improve therapeutic research aims in the field of AD and similar diseases.

**Keywords:** Alzheimer's Disease; Inflammation Mediators; Cytokines; Chemokines; Complement System Proteins; Acute Phase Proteins.

**المخلص:** داء الزهايمر هو انتكاسة عصبية تحدث مع التقدم في العمر وهي أكثر أسباب خرف الشيخوخة شيوعاً. ويتميز المرض بحدوث خلل إدراكي، وترسب لويحات أميلويدية، وتشابك الألياف العصبية مع التهاب عصبي. والسمة المميزة لداء الزهايمر هي حدوث التهاب المخ. وبالتالي فإن الأدلة السريرية والمناعية المرضية قد يتم دعمها بالوسائط الالتهابية، مثل السيتوكينات والكيموكينات، ونظام الكومبليمنت (المتمة)، وعلامات الالتهاب الحاد، والوسائط المؤكسدة. وعلى وجه الخصوص فإن الوسائط المؤكسدة قد يكون لها دور نشط في تفاقم داء الزهايمر والتهاب المخ الجاري. وهذا الاستعراض يقدم لمحة عامة على وظائف وأنشطة وسائط الالتهاب في داء الزهايمر. إن تحسن فهمنا لدور الالتهاب في حدوث داء الزهايمر سيؤدي إلى تقدم البحث العلمي في إمكانية علاج هذا المرض وبالتالي أمراض أخرى مشابهة.

**مفتاح الكلمات:** داء الزهايمر؛ وسائط الالتهاب؛ السيتوكينات؛ الكيموكينات؛ بروتينات نظام الكومبليمنت؛ علامات الالتهاب الحاد.

**A**LZHEIMER'S DISEASE (AD) IS A CHRONIC neurodegenerative disease considered to be the most common cause of dementia.<sup>1</sup> AD is characterised by visuospatial *dysgnosia* and memory, language, emotional, personality and complex cognition impairment. The primary sign of AD is the gradual deterioration in an individual's ability to remember recent events, likely due to the perturbation of neuronal function in the temporal lobes.<sup>1,2</sup> More than 44 million people worldwide were estimated to be suffering from AD in 2014 and this number is projected to double by 2030.<sup>3</sup> There is a direct correlation between the incidence of dementia and ageing, with the highest rates of AD seen in the seventh and eighth decades of life. In addition, it is proposed that the incidence of AD may dramatically rise every five years after the age of 65 years.<sup>4</sup>

The most common neuropathological hallmarks of AD are deposition of amyloid  $\beta$  ( $A\beta$ ) in a compact structure outside the neurons, intracellular neurofibrillary tangles (NFTs) and inflammatory processes.  $A\beta$  is derived from the amyloid precursor protein (APP) by processing enzymes ( $\alpha$ -,  $\beta$ - and  $\gamma$ -secretases). These altered proteins are deposited as extracellular plaques called senile plaques.<sup>5-7</sup> Intracellular NFTs, which are composed of microtubule-associated protein tau, are another pathological aspect of AD-affected brains. Following chemical changes such as hyperphosphorylation, these aggregates often pair with other threads and accumulate inside the neurons, consequently causing microtubule destabilisation.<sup>5,8,9</sup>

Histopathological evidence from patients with AD shows cerebral atrophy, deposition of  $A\beta$  in plaques and neuritic changes, such as neuritic plaques and

<sup>1</sup>Department of Laboratory Medicine, Imam Hassan Mojtaba Hospital, Alborz University of Medical Sciences, Karaj, Iran; <sup>2</sup>Research Center for Immunodeficiencies, Children's Medical Center and <sup>3</sup>Department of Immunology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran; <sup>4</sup>Health & Social Services Sector, The Research Council Oman, Muscat, Oman; <sup>5</sup>Department of Immunology, Faculty of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

\*Corresponding Author e-mail: mirshafiey@tums.ac.ir

**Table 1:** Inflammatory components contributing to the pathology of Alzheimer's disease

Inflammatory component	Examples
CNS cells	Microglia, astrocytes and neurons
Cytokines	TNF- $\alpha$ , IL-1 $\beta$ , IL-6, IFN- $\gamma$ and IL-18
Chemokines	MCP-1, CXCL8, CXCL12, CX3CL1 and MIP
Complement system	Classical and alternative pathways
Acute phase proteins	CRP and SAA
Oxidative mediators	ROS, RNS, NO, O <sub>2</sub> <sup>-</sup> and ONOO <sup>-</sup>

*CNS = central nervous system; TNF- $\alpha$  = tumour necrosis factor- $\alpha$ ; IL = interleukin; IFN- $\gamma$  = interferon- $\gamma$ ; MCP-1 = monocyte chemoattractant protein-1; CXCL = C-X-C motif ligand; CX3CL1 = C-X<sub>3</sub>-C motif ligand 1; MIP = macrophage inflammatory protein; CRP = C-reactive protein; SAA = serum amyloid A; ROS = reactive oxygen species; RNS = reactive nitrogen species; NO = nitric oxide; O<sub>2</sub><sup>-</sup> = superoxide anion radical; ONOO<sup>-</sup> = peroxynitrite anions.*

NFTs. Although the brains of people diagnosed with AD have increased A $\beta$  deposition, these findings are not specific to AD alone and may be found in elderly people not suffering from dementia.<sup>10,11</sup> Although the amyloid cascade theory is the most widely accepted explanation for the aetiology of AD, other theories have also been described to illustrate the role of inflammation in actively contributing to the progression of the disease.<sup>12</sup> It is often assumed that the accumulation of A $\beta$  in the brain results in the development of systemic inflammatory reactions by prompting immune responses.<sup>13,14</sup> This article reviews the major role of inflammatory mediators in the immunopathogenesis of AD.

## Immunopathology of Alzheimer's Disease

Cells of the adaptive immune system can migrate to the brain through the blood-brain barrier (BBB), which is structurally different in people with AD in comparison to healthy individuals.<sup>15</sup> Based on electron microscope observation, most A $\beta$  plaques in the brains of AD patients are associated with activated microglial cells (central nervous system [CNS]-resident macrophages). These cells seem to be responsible for on-going neuro-inflammatory processes in AD through the release of cytokines, chemokines and neurotoxins. Moreover, microglial cells can produce several pro- and anti-inflammatory cytokines via direct inter-action with infiltrated T lymphocytes.<sup>15</sup> Interestingly, Bromley *et al.* reported that some chemokines are able to restrain immunological synapse formation and T cell activation.<sup>16</sup> They indicated that the immunosuppressive effect of chemokines occurred

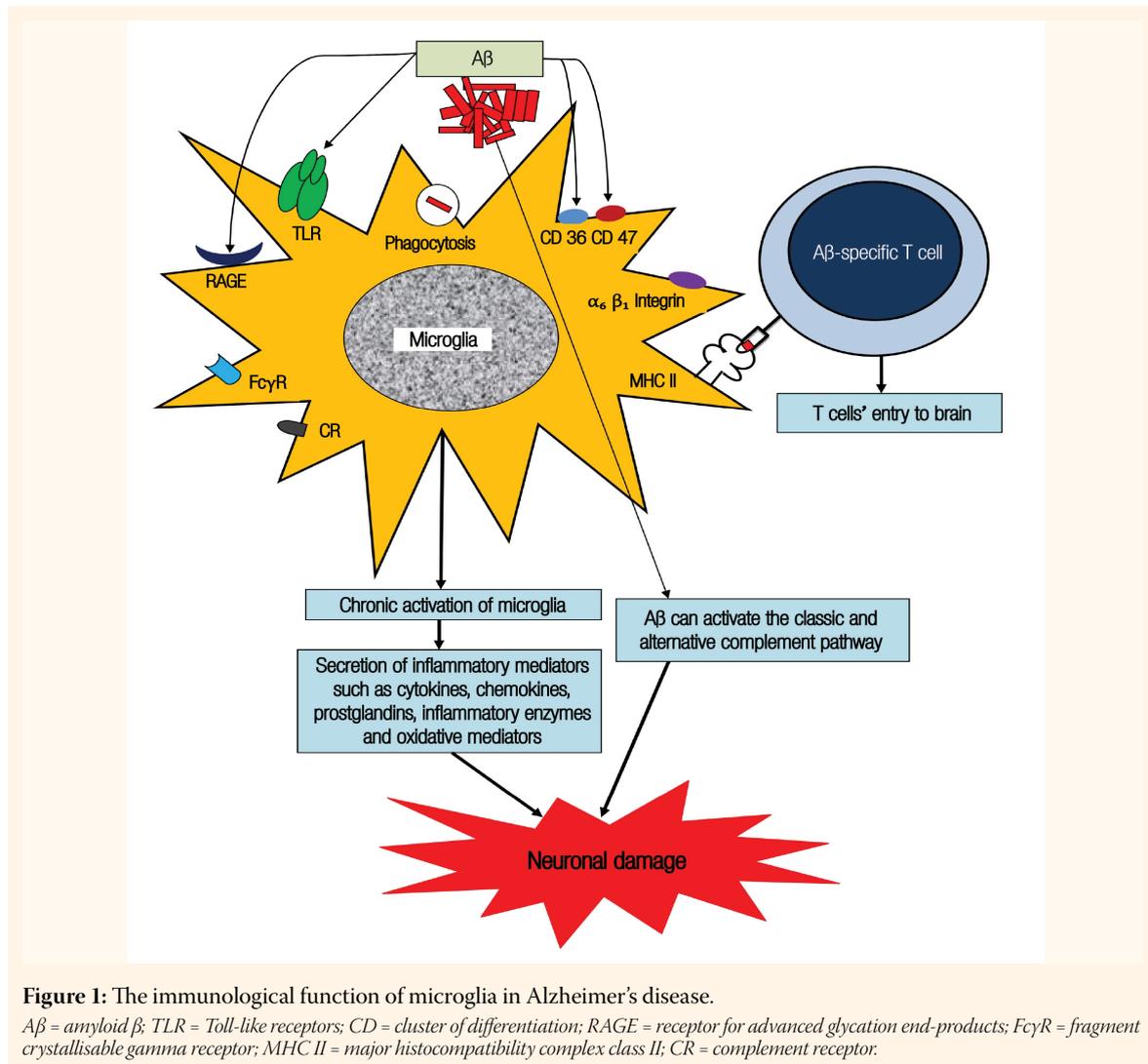
with C-X-C motif receptor (CXCR) 3 and C-C motif receptor (CCR) 7 chemokines, but not with CCR2, 4 and 5 or CXCR4 chemokines.<sup>16</sup>

Inflammation commences when innate immune mediators detect damaged tissue or other molecules on the surface of cells. Chemokines and other inflammatory mediators are responsible for the recruitment of immune cells to the damaged area.<sup>17,18</sup> Various inflammatory processes in AD contribute to the pathology of the disease [Table 1]. Degenerated cells and tissues, as well as an accumulation of abnormal insoluble materials, are the most common stimuli for inflammation. Likewise, in the brain of AD patients, damaged neurons and neurites, along with NFT and insoluble A $\beta$  peptide deposits, can act as potent triggers of inflammation.<sup>19</sup>

Accordingly, inflammation may contribute to the pathogenesis of AD by two mechanisms. The first is a preliminary innate immune response to the alterations in the AD brain; inflammation is involved in the recruitment of immune cells to the site of injury as a result of the initial signalling of cytokines and chemokines and complement system activation. The second mechanism involves a minor amount of on-going inflammation in the brain, which can result in the pathogenesis of AD. This inflammation can be considered a sign of an impaired adaptive immune response and leads to chronic inflammation.<sup>19,20</sup>

## GLIAL CELLS AS A TRIGGER OF IMMUNE RESPONSES

Microglial cells have the ability to induce neuronal damage through the following processes: (1) phagocytosis; (2) the release of cytokines/chemokines/prostaglandins and reactive oxygen species (ROS); and (3) the expression of innate and adaptive immune function molecules such as Toll-like receptors (TLRs), immunoglobulin fragment crystallisable gamma receptors, major histocompatibility complex class II (MHC II) molecules, complement receptors and purinergic receptors (e.g. P2X purinoceptor 7).<sup>21-23</sup> The activation of microglia with A $\beta$  can occur either via the internalisation of soluble A $\beta$  through phagocytosis while fibrillary A $\beta$  binds to TLR2 and TLR4 or through the activation of a mitogen-activated protein kinase pathway, stimulating pro-inflammatory gene expression leading to the secretion of cytokines and chemokines.<sup>24</sup> Moreover, A $\beta$  may be presented by activated microglial cells to T lymphocytes, eventually causing A $\beta$ -specific T cells to enter the brain [Figure 1]. It should be noted that while A $\beta$ -reactive T cells are present in healthy individuals, T cells are vital for protecting against AD pathogenesis by cooperating with and modulating the innate immune system.<sup>25</sup>



However, T cells are more susceptible to ageing than innate immune components. Fulop *et al.* demonstrated that adaptive immune responses in the elderly were less effective in eliminating Aβ deposition than an innate immune response.<sup>25</sup> Therefore, once the immune system is overwhelmed with Aβ deposition, inflammation becomes chronic and adverse, resulting in accelerated neurodegeneration.<sup>26</sup>

### ROLE OF LYMPHOCYTES

Several studies have evaluated the function of T cells in AD so as to determine AD-related abnormalities in the immune system.<sup>27,28</sup> Richartz-Salzburger *et al.* reported the general decline of immune function among patients with AD due to a decreased number of T and B cells.<sup>27</sup> In addition, another study noted the hyporesponsiveness of T cells to certain intrinsic functional defects in AD patients when compared with control subjects.<sup>29</sup> Some alterations, such as accelerated telomere shortening, can serve as an important factor in the impairment of normal lymphocyte

activity in AD patients.<sup>30</sup> This finding was confirmed by Zhang *et al.*; their research indicated that the increase of telomerase activity in the lymphocytes of AD patients can lead to diminishing lymphocyte proliferation activity, consequently resulting in the loss of immune function.<sup>31</sup>

As yet, the precise mechanism regarding the activation, migration and survival of T cells in the brains of AD patients is not clear. As previously mentioned, antigen-presenting cells with a high expression of MHC II molecules, which either differentiate from microglia in the brain or are recruited from the blood, can present Aβ to T cells. In addition, interferon-γ (IFN-γ) plays a key role in facilitating T cell migration as well as promoting an immune regulatory process within the brain.<sup>32</sup> The amount of IFN-γ in the brain determines its effect as it is adverse at high levels and beneficial at low levels.<sup>33</sup> Research has demonstrated the pathogenic reaction of both T and B cells against Aβ as well as the risk of meningoencephalitis, caused by the entry of

cluster of differentiation (CD) 8 cytotoxic T cells to the brain followed by the secretion of pro-inflammatory cytokines by CD4 T cells.<sup>33,34</sup>

### ROLE OF INFLAMMATORY CYTOKINES AND CHEMOKINES

There is growing evidence that inflammatory mediators in the CNS contribute to cognitive impairment through cytokine-mediated interactions between glial cells and neurons. Moreover, it has been demonstrated that AD is associated with the upregulation of pro-inflammatory cytokines, which can initiate plaque production and enhance nerve cell degeneration.<sup>6,15</sup> Some of these mediators—including tumour necrosis factor (TNF)- $\alpha$ , interleukin (IL)-6, IFN- $\gamma$ , inducible protein-10, monocyte chemoattractant protein (MCP)-1 and C-X-C motif ligand (CXCL) 8—increase in the prodromal stage of AD. Elevation of these cytokines occurs during the initiation of AD. This may explain the failure of clinical trials using anti-inflammatory drugs against severe AD.<sup>35</sup> This idea is supported by immunohistochemistry examinations performed by Sokolova *et al.* which confirmed the increase of MCP-1, IL-6 and CXCL8 in AD brains and determined that these mediators were localised in neurons (IL-6, MCP-1 and CXCL8), astrocytes (IL-6 and MCP-1) and plaques (CXCL8 and MCP-1).<sup>36</sup> Moreover, logistic linear regression modelling determined that, of the cytokines, MCP-1 was the most accurate for the prediction of AD.<sup>36</sup> Hence, these findings support the importance of IL-6, MCP-1 and CXCL8 in AD and also show that MCP-1 may play an important role in the progression of chronic inflammation in AD. As a result of these observations, some cytokines (such as IL-1, -4, -6, -10, -12 and -18, IFN- $\gamma$ , TNF and transforming growth factor [TGF]- $\beta$ ) have been proposed as AD biomarkers.<sup>6</sup>

#### ***Interleukin (IL)-1 $\beta$ , IL-6 and Tumour Necrosis Factor- $\alpha$***

The elevation of IL-1 $\beta$ , IL-6 and TNF- $\alpha$  is widely recognised as a critical component of neuro-inflammation and leukocyte recruitment to the CNS.<sup>6</sup> This response is characterised by promoting deposition of A $\beta$  in the brain and astrocytic and microglial activation. Moreover, IL-1 $\beta$  and TNF- $\alpha$  are potent stimuli for inducible nitric oxide (NO) synthase (NOS) expression and activity in the brain and NO metabolite overflow into the cerebrospinal fluid.<sup>5</sup> Importantly, Belkhef *et al.* revealed recently that a high level of NO is associated with the rise of TNF- $\alpha$  levels in patients in the severe stages of AD.<sup>37</sup> In response to numerous intrinsic and extrinsic stimuli, TNF- $\alpha$  is produced by microglia, astrocytes and neurons in the brain. In addition, genetic and

epidemiological findings have implicated augmented levels of TNF- $\alpha$  in the brain as a risk factor for AD.<sup>37</sup>

TNF- $\alpha$  can mediate neuronal dysfunction as well as A $\beta$ -induced disruption of the molecular mechanisms involved in memory function. Likewise, TNF- $\alpha$  can stimulate accumulation of the tau proteins in neurites through induction of ROS.<sup>5</sup> In another recent study, Lin *et al.* observed a significant decrease of TNF- $\alpha$ , IL-1 $\alpha$ , -6 and -12 in sera after vaccinating transgenic mice.<sup>38</sup> Remarkably, the decrease in TNF- $\alpha$  and IL-6 levels correlated with cognitive and behavioural improvements in the transgenic mouse model of AD.<sup>38</sup> In contrast, it has been reported that some inflammatory cytokines, such as IL-1 $\beta$ , IL-6 and IFN- $\gamma$ , have also had beneficial and protective effects against AD.<sup>6,39</sup> Overexpression of IL-1 and IL-6 in the brain results in extensive *gliosis* which may be beneficial in the disease process by stimulating increased amyloid phagocytosis rather than mediating a neurotoxic feedback loop.

#### ***Interleukin-18 and Interferon- $\gamma$***

Several studies have highlighted a critical role for IL-18 in mediating neuro-inflammation and neurodegeneration in the brains of AD patients.<sup>40,41</sup> Notably, an imbalance of IL-18 and its endogenous inhibitor, IL-18 binding protein (IL-18BP), has been shown in AD, with an elevated IL-18:IL-18BP ratio that might be involved in the disease immunopathogenesis.<sup>42</sup> In the brain, IL-18 is produced by microglial, astrocyte and ependymal cells as well as by neurons of the medial habenular nucleus.<sup>43</sup> Through the induction of IFN- $\gamma$  and expression of MHC II molecules in microglial cells, IL-18 can initiate a neural-immune cell interaction which may play a key role in the induction of autoimmunity in the CNS environment.<sup>44</sup> Moreover, IFN- $\gamma$  can enhance A $\beta$  deposition through  $\beta$ -secretase 1 expression as well as stimulate the upregulation of MHC II molecules in a subpopulation of microglia and induce auto-immune processes in the CNS. Importantly, significant IFN- $\gamma$  levels are only detected in mild cases of AD. Collectively, this suggests that NO production is IFN- $\gamma$ -dependent in AD.<sup>37</sup> On the other hand, IFN- $\gamma$  is a known inhibitor of APP fragment production. IFN- $\gamma$  can prevent amyloid deposition during inflammatory processes in both non-neuronal and neuronal tissues. Furthermore, IFN- $\gamma$  has a strong suppressive effect on the production and metabolism of APP.<sup>5,45</sup>

#### ***Transforming Growth Factor- $\beta$***

Recent data have implicated anti-inflammatory cytokines as integral factors to the pathogenesis of AD.<sup>6</sup> Among them, TGF- $\beta$  is emerging as a critical factor in regulating inflammatory responses. Three known

isoforms of TGF- $\beta$  (TGF- $\beta$ 1, - $\beta$ 2 and - $\beta$ 3) are expressed in mammalian tissues. In AD, the expression of TGF- $\beta$ 2 is induced by toxic A $\beta$  in both glial and neuronal cells; increased levels of TGF- $\beta$ 2 trigger the cell death pathway due to alterations in the morphology and number of lysosomes in neurons.<sup>46</sup> TGF- $\beta$ 2 causes lysosomal membranes to become unstable and leak and this effect is intensified with the accumulation of A $\beta$ , as TGF- $\beta$ 2 rapidly targets the A $\beta$  peptide in lysosomal compartments in cortical neurons, inducing cell death.<sup>47</sup> As a neurotrophic factor, TGF- $\beta$ 1 initiates and maintains neuronal differentiation and synaptic plasticity. In AD animal models, it has been suggested that a deficiency of TGF- $\beta$ 1 signalling may correlate with A $\beta$  pathology and NFT formation.<sup>48</sup>

#### **C-C Motif Ligand 2**

C-C motif ligand (CCL) 2, also known as MCP-1, plays a significant role in AD pathogenesis. Increased levels of CCL2 in the brain result in the recruitment of activated monocyte cells into the organ where they differentiate into macrophages and produce neurotoxic and inflammatory mediators.<sup>49–51</sup> Immunohistochemistry findings have confirmed this increase in CCL2 levels and have determined localisation of these factors in astrocytes, neurons and plaques via pathology.<sup>49</sup> However, mononuclear phagocyte accumulation is regulated via the interaction of CCL2 with its receptor, CCR2; CCR2 deficiency in these cells therefore leads to diminished phagocyte cell recruitment to the brain which is associated with higher levels of A $\beta$  in the brain, particularly around the blood vessels. This suggests that monocytes are initially recruited and accumulate at A $\beta$  deposition sites in order to clear them and either halt or delay their associated neurotoxic effects. Indeed, an increase of mononuclear phagocyte recruitment to the brain delays the progression of AD in its early stages.<sup>36,49</sup>

#### **C-C Motif Ligand 5**

The role of CCL5, also known as the RANTES (regulates on activation, normal T cell expressed and secreted) protein, has been determined in neurodegenerative diseases such as AD and elevated levels of RANTES protein are commonly observed in the microcirculatory system of AD-affected brains.<sup>52</sup> The RANTES protein, as well as several other chemokines in astrocytes, is upregulated as a response to a cytokine-mediated increase of ROS.<sup>53</sup> Moreover, oxidative stress upregulates RANTES protein expression in endothelial cells in the brain.<sup>52</sup> In brain injury models, elevated levels of the RANTES protein contributed to immune cell recruitment that occurred concurrently with increased rates of neuronal death.<sup>52,54,55</sup>

#### **C-X-C Motif Ligand 8**

CXCL8 (IL-8) is a microglia-derived chemokine that is produced in response to pro-inflammatory signals such as A $\beta$ . CXCL8 could be important for the recruitment of activated microglia and neutrophils into areas of the damaged brain during the late stages of AD, suggesting a role for this chemokine in phases with prevalent neurodegeneration. In addition, CXCL8 is continually upregulated in neurons and plaques.<sup>19</sup>

#### **C-X-C Motif Ligand 12**

The chemokine CXCL12 has been associated with neurogenesis and the recruitment of brain-resident and non-resident circulating cells to sites of lesions in the CNS.<sup>56</sup> Moreover, in Tg2576 mouse models of AD, CXCL12 messenger ribonucleic acid (mRNA) protein and its receptors were downregulated, with co-existing cognitive deficits.<sup>57</sup> Zhu *et al.* found that CXCL12 plasma levels in patients with early AD were low and that CXCR4 and CXCL12 had anti-inflammatory properties.<sup>58</sup> An *in vitro* study demonstrated that neurons pre-treated with CXCL12 were significantly protected from antibody-induced dendritic regression and *apoptosis* via protein kinase B and extracellular signal-regulated kinases 1/2 activation, as well as maintenance of A disintegrin and metalloproteinase 17, especially with CXCL12.<sup>59</sup>

#### **C-X3-C Motif Ligand 1**

C-X3-C motif ligand (CX3CL) 1, also known as fractalkine, is produced and expressed constitutively by neurons. CX3CL1 suppresses microglial activation and the CX3CL1/C-X3-C motif receptor (CX3CR) 1 complex may control neurotoxicity. Research has demonstrated that levels of plasma-soluble CX3CL1 are significantly greater in patients with mild to moderate AD than in those with severe AD.<sup>60</sup> These findings and other data suggest that CX3CL1 has a neuroprotective function that may potentially have therapeutic applications for several neurodegenerative diseases, including AD and Parkinson's disease, in which inflammation also plays an important role.<sup>5,60</sup> Cho *et al.* identified CX3CL1/CX3CR1 signalling as a central microglial pathway in protecting against AD; this pathway was associated with the inhibition of aberrant microglial activation and inflammatory cytokine elevation.<sup>61</sup>

#### **Macrophage Migration Inhibitory Factor**

The macrophage migration inhibitory factor (MIF) is a pleiotropic pro-inflammatory cytokine which increases the production of other inflammatory cytokines, such as TNF- $\alpha$ , IL-6 and IFN- $\gamma$ , and has a pivotal regulatory role in the pathogenesis of numerous autoimmune and inflammatory disorders.<sup>62,63</sup> Cellular

sources for MIF are neurons and activated microglia; the clustering of microglia at amyloid deposition sites implies that this cell migrates to these locations and undergoes attempted removal of the amyloid protein.<sup>64</sup> In their study, Oyama *et al.* found that MIF was able to bind A $\beta$  in AD-affected brains and that A $\beta$  toxicity could thus be accredited directly to the increased expression of MIF.<sup>65</sup> However, microglial cells are seemingly unable to clear A $\beta$  due to its insoluble nature and the fact that it is present in substantial quantities. Therefore, A $\beta$  deposits are not phagocytised and remain present while microglial cells continue to be attracted to the sites for long periods of time.<sup>66</sup>

### **Macrophage Inflammatory Protein-1 $\alpha$**

Macrophage inflammatory protein (MIP)-1 $\alpha$  is a chemokine present in humans which has a significant part in the pathogenesis of AD mainly via its expression by astrocytes, microglia, neurons, infiltrated monocytes and T cells.<sup>67</sup> It has been demonstrated that higher MIP-1 $\alpha$  expression in the peripheral T lymphocytes of AD patients results in CCR5 expression, a potential MIP-1 $\alpha$  receptor on microvascular endothelial cells in the human brain, which subsequently leads to increased T cell transendothelial migration from the blood to the brain.<sup>68</sup> Li *et al.* found that serum MIP-1 $\alpha$  levels were significantly higher in patients with the TA6/6 genotype of the *apolipoprotein E* gene and that this genotype seems to be a genetic risk factor for AD.<sup>69</sup> According to Passos *et al.*, the activation of the MIP-1 $\alpha$ /CCR5 signalling pathway was one of the initial events following A $\beta$ 1–40 injections in AD mouse models; this seems to be a critical signal for activated glial cell accumulation, inflammatory responses, synaptic dysfunction and cognitive failure.<sup>70</sup>

### **CONTRIBUTION OF THE COMPLEMENT SYSTEM**

In the AD-affected brain, levels of complement mRNAs and their protein products have been found to be significantly higher than those in the livers of healthy individuals; moreover, neurons of AD patients have been found to express complement proteins of the classical pathway to an increased degree in comparison to neurons in brains which were not affected by AD.<sup>9</sup> At the very first stages of amyloid deposition in AD, integral complement protein components of amyloid plaques and cerebral vascular amyloid material can be found; their activation occurs simultaneously with the clinical expression of AD.<sup>71</sup> A $\beta$  can activate the classic and alternative complement pathways in areas of the brain associated with AD pathology, even in the absence of antibodies.<sup>72</sup> Moreover, it has been demonstrated that tau protein is an antibody-independent activator of

the classical complement pathway.<sup>9</sup> This activation of the complement cascade not only causes substantial damage to the neurons, but can also lead to increased phosphorylation of tau proteins and formation of NFTs, resulting in elevated levels of membrane attack complex in AD brains.<sup>9</sup> Nevertheless, the complement cascade has both positive and negative aspects; although it is essential to maintaining the health of the brain, it may have adverse effects when unregulated and often exacerbates AD.

The complement protein C1q is induced in the brain in response to AD and blocks fibrillary A $\beta$  neurotoxicity *in vitro*.<sup>73</sup> C1q binds to  $\beta$ -sheet fibrillary A $\beta$  plaques and, when associated with C1r and C1s as in the C1 complex, activates the complement cascade that can have detrimental inflammatory consequences via production of the chemotactic factor C5a and following recruitment and activation of microglial cells to the site of injury. It also has a protective effect by increasing the clearance of A $\beta$  through C1q- and C3-dependent opsonisation.<sup>74</sup> Benoit *et al.* showed that C1q protected both immature and mature primary neurons against fibrillary A $\beta$  toxicity and prevented oligomeric A $\beta$  toxicity.<sup>75</sup> In addition, gene expression analysis revealed that C1q-induced low-density lipoprotein receptor-related protein 1B and G protein-coupled receptor 6 expressed early in AD mouse models were vital for C1q-mediated protection against A $\beta$  neurotoxicity.<sup>75,76</sup> Loeffler *et al.* noted that C3b and iC3b (a cleavage product of C3b) are also deposited on AD-affected neurons, much like C1q.<sup>77</sup>

### **FUNCTION OF ACUTE PHASE PROTEINS**

Several prospective epidemiological studies have found that increased acute phase mediator serum levels can serve as a risk factor for AD, as detailed in a review article by Eikelenboom *et al.*<sup>78</sup> In addition, other clinical studies compiled in the review have suggested that increased peripheral inflammation is associated with a greater risk of dementia, mainly in patients with pre-existing cognitive impairments, and accelerates subsequent deterioration in patients with dementia.<sup>78</sup> Higher levels of serum C-reactive protein (CRP) in middle-aged patients are also associated with an increased risk of AD and vascular dementia, which may indicate that inflammatory factors are a reflection of both dementia-related peripheral disease and cerebral mechanisms.<sup>78</sup> Of note is the fact that these processes can be measured long before manifestations of dementia begin to be observed. Follow-up studies in the elderly have also revealed a correlation between serum CRP levels and an increased incidence of dementia and AD.<sup>4</sup> Kravitz *et al.* reported that high CRP levels were related to the increased likelihood of

all-cause dementia occurring in the elderly, particularly for females.<sup>79</sup> Moreover, Komulainen *et al.* found that elevated high-sensitivity CRP (hsCRP) serum concentrations were a predictor of poorer memory function in women 12 years after the measurements had been taken.<sup>80</sup> Therefore, hsCRP may be a useful biomarker to identify individuals with an increased risk for cognitive decline.

Strang *et al.* discussed the fact that no pathomechanistic link has yet been established between circulating pentameric CRP (pCRP) and AD, despite reports of an association between the two.<sup>81</sup> Their hypothesis was that A $\beta$  plaques induce the dissociation of pCRP to single monomers, which have more potent pro-inflammatory properties than pCRP, and the inflammation is subsequently localised to the AD plaques.<sup>81</sup> Helmy *et al.* presented evidence that serum levels of IL-6 and CRP were significantly elevated among patients with vascular and Alzheimer's dementia in comparison to elderly subjects in good health.<sup>82</sup> Although IL-6 levels were higher in AD patients in comparison to those with vascular dementia, the difference was not found to be significant. Furthermore,  $\alpha_1$ - and  $\alpha_2$ -globulins were significantly higher in AD patients and researchers were able to distinguish vascular from Alzheimer's dementia.<sup>82</sup>

Elevations in CRP in middle-aged patients have been associated with an increased risk of AD development. O'Bryant *et al.* reported decreased CRP levels in AD patients; in fact, mean CRP levels were found to be significantly reduced in AD patients versus controls (2.9 versus 4.9  $\mu\text{g/mL}$ , respectively).<sup>83</sup> However, Sundelöf *et al.* reported contradictory findings which indicated that hsCRP and serum amyloid A levels were not associated with AD risk in elderly men.<sup>84</sup>

## OXIDATIVE STRESS

Cellular oxidative stress—including enhanced protein oxidation and nitration, glycoloxidation, lipid peroxidation and A $\beta$  accumulation—is associated with AD.<sup>85</sup> The deposition of A $\beta$  generates ROS, which is involved in the inflammatory and neurodegenerative pathology of AD. Oxidative stress can therefore exacerbate the progression of AD. When repair attempts are made by the brain to remedy oxidative damage, characterised by APP overexpression, adenosine triphosphate (ATP)-binding cassette sub-family G member 2 (ABCG2) is upregulated and activator protein-1 is activated. Not only do these proteins stop blood A $\beta$  from entering the brain via the BBB but they also protect against oxidative stress by decreasing ROS production, boosting antioxidant activity and inhibiting the inflammatory response through the inhibition of the nuclear factor- $\kappa\text{B}$  signalling pathway in brain tissue. As

a result, ABCG2 may have a protective function in the neuroinflammatory response of AD.<sup>85–87</sup> Additionally, the apparent end-product of APP, the formation and accumulation of A $\beta$ , appears to be initiated by ABCG2.

This process can lead to increased free radical production—mainly superoxide anions via the mitochondria—which induces the interruption of oxidative phosphorylation and engenders a decrease in ATP levels. The mitochondrial dysfunction and damage that occurs with ageing correlates with the augmented intracellular production of oxidants and pro-oxidants. The extended oxidative stress in brain tissue, and the resultant hypoperfusion, stimulates the expression of NOS which subsequently drives the formation of ROS and reactive nitrogen species (RNS). ROS contributes to the dysfunction of the BBB and damage to the brain's parenchymal cells. Moreover, it has been shown that ROS is potentially toxic and may damage the proteins, lipids and nucleic acids of brain cells and mitochondria, including neurons and oligodendrocytes that may mediate toxicity.<sup>88,89</sup> Generation of ROS is controlled by sensitive genes called vitagenes. These genes encode proteins such as heat shock proteins, nutritional antioxidants which play a neuroprotective role.<sup>90</sup>

Oxidative stress could also lead to further damage in AD-affected brains through inducible NOS overexpression and constitutive neuronal NOS activity, which increases the production of NO and its derivative RNS. In an AD-affected brain, NO and superoxide anion radicals (O $_2^-$ ) are produced by reactive astrocytes and microglia in response to A $\beta$ .<sup>91–93</sup> Formed from NO and O $_2^-$ , peroxynitrite anions are another component of oxidative stress. These extremely reactive oxidising and nitrating agents lead to the oxidation of cellular components, increased A $\beta$  aggregation and a stimulated inflammatory response.<sup>94</sup>

In the early stages of AD, the pathology shows that inducible neuron-specific cyclooxygenase-2 (COX-2) enzymes are expressed and upregulated by neuronal cells which are closely linked with the A $\beta$ -bearing cells. It has been suggested that A $\beta$  can stimulate activity of COX-2 oxygenase and peroxidase in a cell-free system—this stimulation of the two-step action of COX-2 leads to the production of ROS and prostaglandin E $_2$ .<sup>85,95–97</sup> In addition, recent findings in mouse models suggest a role for COX/prostaglandin E $_2$  signalling in the development of AD.<sup>98</sup>

## Treatment and Immunotherapy for Alzheimer's Disease

There is currently no cure available for AD; however, drug and non-drug treatments may help with both

cognitive and behavioural symptoms of the disease. Two types of medications designed to treat the cognitive manifestations of AD have been approved by the Food and Drug Administration in the USA: cholinesterase inhibitors and a new N-methyl-D-aspartate receptor antagonist, memantine.<sup>99</sup> Three forms of cholinesterase inhibitor drugs are commonly prescribed: donepezil (approved to treat all stages of AD), rivastigmine and galantamine (both approved to treat only mild to moderate cases of AD).<sup>100</sup>

Inflammation is one of numerous hypotheses that have been proposed for the multifactorial aetiology of AD; indeed, inflammation may interact with other triggers in several ways. This network of mechanisms makes it difficult to identify any specific inflammatory process, causal factor or cell in order to determine their individual role in AD.<sup>101</sup> Risk factors for AD, which may include genetic, biological and environmental factors, contribute to neuro-inflammation and to subsequent neurodegeneration in the later stages of AD. However, they may have fewer effects on the early pathogenesis of the disease.<sup>102,103</sup> Therefore, due to the distinctive role of inflammation in the early versus late stages of AD, anti-inflammatory agents, such as non-steroidal anti-inflammatory drugs (NSAIDs), may potentially be a treatment option for AD patients, although this would be dependent on the stage of the disease.

Researchers have reported a decrease in AD development among subjects taking NSAIDs for long periods of time; thus, it has been proposed that NSAIDs could directly reduce the production of A $\beta$  through several mechanisms.<sup>9,104</sup> Unfortunately, conflicting results have been reported in the literature and related clinical trials have not yet yielded promising findings. The toxic effects of NSAID treatments also prevent their widespread use.<sup>9,104</sup> Additionally, anti-inflammatory medications may have no effect on patients in later stages of the disease. This is because the most important aetiological factors for early-onset AD are the mismetabolism of APP along with the increased production of A $\beta$  followed by the deposition of fibrillary A $\beta$ , which can activate the innate immunity receptors leading to activation of microglia and reactive astrocytes. This exacerbates neurodegeneration through the release of inflammatory cytokines, ROS and other factors. Modifications of these factors can occur very early during the development of the disease; trials with anti-inflammatory agents may therefore be ineffective in patients with severe AD.<sup>6</sup> Furthermore, the efficacy of anti-inflammatory drugs such as aspirin, steroids and other traditional NSAIDs and COX-2 inhibitors in AD patients has not yet been proven; thus, these drugs cannot be recommended for AD treatment.<sup>105</sup>

Immunotherapy has been proposed as a potential candidate for the treatment of AD. Both active and passive A $\beta$  immunotherapies have been developed to decrease the load of A $\beta$  by enhancing its rate of elimination. Vaccinations, in the form of active immunisation with A $\beta_{42}$  (the common form of A $\beta$  in amyloid plaques) or other synthetic peptides, have been successfully assessed in transgenic animal models of AD.<sup>106</sup> The basis of this approach is the priming of T, B and microglial cells, which provoke immune responses.

One type of passive immunotherapy, administering monoclonal antibodies against the A $\beta$  fragment, diminishes the need for patients to mount immunity against A $\beta$  peptides. An on-going clinical trial in the USA sponsored by a pharmaceutical company (Eli Lilly & Co., Indianapolis, Indiana, USA) is currently testing to see if treatment with solanezumab, a monoclonal antibody against A $\beta$ , significantly slows the loss of awareness and cognitive and functional decline in patients with mild AD.<sup>107</sup> However, concerns exist regarding the use of related monoclonal antibodies as a therapeutic option. Firstly, new approaches are needed due to the poor penetration of antibodies into the brain and, secondly, recognition of the clearance pathways of A $\beta$ /anti-A $\beta$  immune complexes is essential to circumvent obstruction of these pathways during treatment.<sup>106</sup>

## Recent Advances in Alzheimer's Disease

A prospective longitudinal study by Bateman *et al.* has indicated that A $\beta$  deposition in the brain is detectable more than 20 years prior to the onset of AD symptoms.<sup>108</sup> In addition, although the production of A $\beta$  in AD patients is similar to that of cognitively normal individuals, clearance of A $\beta$  in the brain of AD patients is significantly reduced in comparison to control subjects.<sup>109</sup> Immunotherapy and the involvement of antibodies could therefore be a successful approach to facilitating this clearance process. Active immunisation with the DNA A $\beta_{42}$  vaccination may be effective in accomplishing this; the method involves injecting DNA encoding A $\beta_{42}$  where it is subsequently translated in the immunised individual to express A $\beta$  peptide which then stimulates the respective immune responses against A $\beta_{42}$ . Qu *et al.* found a 50% reduction in the level of A $\beta_{42}$  plaques in transgenic mouse models when using this approach; this reduction was later confirmed by another study.<sup>110,111</sup>

Currently, other approaches for AD therapy focus on clearance of A $\beta$  fragments by different pathways, including chaperone-mediated and autophagocytic

clearance.<sup>112</sup> Chaperones are a specific cluster of proteins which can correct or prevent the misfolding of proteins. Autophagy is a normal cellular process in the body which preserves homeostasis or normal functioning through protein destruction and turnover of destroyed cell organelles for new cell formation. Several studies have noted the occurrence of autophagocytic vacuoles in the brains of patients with AD.<sup>113,114</sup> Caccamo *et al.* reported that the level of beclin 1, a protein involved in the formation of the autophagosome, is diminished in the brains of AD patients.<sup>115</sup> Moreover, Martorana *et al.* found an augmented subset of B cells with a memory double-negative phenotype in elderly people.<sup>116</sup> Interestingly, Colonna-Romano *et al.* reported that B cells are late memory or exhausted cells, which may be a manifestation of ageing or a dysregulation of the immune system.<sup>117</sup>

## Conclusion

Numerous hypotheses have been proposed for the multifactorial aetiology of AD, including inflammation. Current evidence supports the potential role of inflammation in AD, although this factor may interact with other genetic, biological and environmental triggers in several ways. Immunotherapy and the use of antibodies could have applications for patients with AD. In order to improve the range and efficacy of therapeutic options for AD patients and those with similar neurodegenerative disorders, further research is recommended to advance the current knowledge of inflammatory processes with regards to this form of dementia.

## References

- Alzheimer's Association. 2012 Alzheimer's disease facts and figures. *Alzheimers Dement* 2012; 8:131–68. doi: 10.1016/j.jalz.2012.02.001.
- Oda H, Yamamoto Y, Maeda K. The neuropsychological profile in dementia with Lewy bodies and Alzheimer's disease. *Int J Geriatr Psychiatry* 2009; 24:125–31. doi: 10.1002/gps.2078.
- Alzheimer's Disease International. World Alzheimer report 2014: Dementia and risk reduction - An analysis of protective and modifiable factors. From: [www.alz.co.uk/research/WorldAlzheimerReport2014.pdf](http://www.alz.co.uk/research/WorldAlzheimerReport2014.pdf) Accessed: Feb 2015.
- Qiu C, Kivipelto M, von Strauss E. Epidemiology of Alzheimer's disease: Occurrence, determinants, and strategies toward intervention. *Dialogues Clin Neurosci* 2009; 11:111–28.
- Gorlovoy P, Larionov S, Pham TT, Neumann H. Accumulation of tau induced in neurites by microglial proinflammatory mediators. *FASEB J* 2009; 23:2502–13. doi: 10.1096/fj.08-123877.
- Azizi G, Mirshafiey A. The potential role of proinflammatory and antiinflammatory cytokines in Alzheimer disease pathogenesis. *Immunopharmacol Immunotoxicol* 2012; 34:881–95. doi: 10.3109/08923973.2012.705292.
- Mirshafiey A, Asghari B, Ghalamfarsa G, Jadidi-Niaragh F, Azizi G. The significance of matrix metalloproteinases in the immunopathogenesis and treatment of multiple sclerosis. *Sultan Qaboos Univ Med J* 2014; 14:e13–25.
- Reitz C, Brayne C, Mayeux R. Epidemiology of Alzheimer disease. *Nat Rev Neurol* 2011; 7:137–52. doi: 10.1038/nrneurol.2011.2.
- Tuppo EE, Arias HR. The role of inflammation in Alzheimer's disease. *Int J Biochem Cell Biol* 2005; 37:289–305. doi: 10.1016/j.biocel.2004.07.009.
- Serrano-Pozo A, Frosch MP, Masliah E, Hyman BT. Neuropathological alterations in Alzheimer disease. *Cold Spring Harb Perspect Med* 2011; 1:a006189. doi: 10.1101/cshperspect.a006189.
- Jellinger KA, Bancher C. Neuropathology of Alzheimer's disease: A critical update. *J Neural Transm Suppl* 1998; 54:77–95. doi: 10.1007/978-3-7091-7508-8\_8.
- McGeer PL, McGeer EG. The amyloid cascade-inflammatory hypothesis of Alzheimer disease: Implications for therapy. *Acta Neuropathol* 2013; 126:479–97. doi: 10.1007/s00401-013-1177-7.
- Orsini F, De Blasio D, Zangari R, Zanier ER, De Simoni MG. Versatility of the complement system in neuroinflammation, neurodegeneration and brain homeostasis. *Front Cell Neurosci* 2014; 8:380. doi: 10.3389/fncel.2014.00380.
- Blach-Olszewska Z, Leszek J. Mechanisms of over-activated innate immune system regulation in autoimmune and neurodegenerative disorders. *Neuropsychiatr Dis Treat* 2007; 3:365–72.
- Azizi G, Khannazer N, Mirshafiey A. The potential role of chemokines in Alzheimer's disease pathogenesis. *Am J Alzheimers Dis Other Demen* 2014; 29:415–25. doi: 10.1177/1533317513518651.
- Bromley SK, Peterson DA, Gunn MD, Dustin ML. Cutting edge: Hierarchy of chemokine receptor and TCR signals regulating T cell migration and proliferation. *J Immunol* 2000; 165:15–19. doi: 10.4049/jimmunol.165.1.15.
- Heneka MT, O'Banion MK, Terwel D, Kummer MP. Neuroinflammatory processes in Alzheimer's disease. *J Neural Transm* 2010; 117:919–47. doi: 10.1007/s00702-010-0438-z.
- Rubio-Perez JM, Morillas-Ruiz JM. A review: Inflammatory process in Alzheimer's disease, role of cytokines. *ScientificWorldJournal* 2012; 2012:756357. doi: 10.1100/2012/756357.
- Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, et al. Inflammation and Alzheimer's disease. *Neurobiol Aging* 2000; 21:383–421. doi: 10.1016/S0197-4580(00)00124-X.
- Zotova E, Nicoll JA, Kalaria R, Holmes C, Boche D. Inflammation in Alzheimer's disease: Relevance to pathogenesis and therapy. *Alzheimers Res Ther* 2010; 2:1. doi: 10.1186/alzrt24.
- Brown GC, Neher JJ. Microglial phagocytosis of live neurons. *Nat Rev Neurosci* 2014; 15:209–16. doi: 10.1038/nrn3710.
- Li XQ, Wang J, Fang B, Tan WF, Ma H. Intrathecal antagonism of microglial TLR4 reduces inflammatory damage to blood-spinal cord barrier following ischemia/reperfusion injury in rats. *Mol Brain* 2014; 7:28. doi: 10.1186/1756-6606-7-28.
- Lee HG, Won SM, Gwag BJ, Lee YB. Microglial P2X7 receptor expression is accompanied by neuronal damage in the cerebral cortex of the APPswe/PS1dE9 mouse model of Alzheimer's disease. *Exp Mol Med* 2011; 43:7–14. doi: 10.3858/emmm.2011.43.1.001.
- Ho GJ, Drego R, Hakimian E, Masliah E. Mechanisms of cell signaling and inflammation in Alzheimer's disease. *Curr Drug Targets Inflamm Allergy* 2005; 4:247–56. doi: 10.2174/1568010053586237.
- Fulop T, Lacombe G, Cunnane S, Le Page A, Dupuis G, Frost EH, et al. Elusive Alzheimer's disease: Can immune signatures help our understanding of this challenging disease? Part 2: New immune paradigm. *Discov Med* 2013; 15:33–42.

26. Querfurth HW, LaFerla FM. Alzheimer's disease. *N Engl J Med* 2010; 362:329–44. doi: 10.1056/NEJMra0909142.
27. Richartz-Salzburger E, Batra A, Stransky E, Laske C, Kohler N, Bartels M, et al. Altered lymphocyte distribution in Alzheimer's disease. *J Psychiatr Res* 2007; 41:174–8. doi: 10.1016/j.jpsychores.2006.01.010.
28. Skias D, Bania M, Reder AT, Luchins D, Antel JP. Senile dementia of Alzheimer's type (SDAT): Reduced T8+-cell-mediated suppressor activity. *Neurology* 1985; 35:1635–8. doi: 10.1212/WNL.35.11.1635.
29. Giubilei F, Antonini G, Montesperelli C, Sepe-Monti M, Cannoni S, Pichi A, et al. T cell response to amyloid-beta and to mitochondrial antigens in Alzheimer's disease. *Dement Geriatr Cogn Disord* 2003; 16:35–8. doi: 10.1159/000069991.
30. Panossian LA, Porter VR, Valenzuela HF, Zhu X, Reback E, Masterman D, et al. Telomere shortening in T cells correlates with Alzheimer's disease status. *Neurobiol Aging* 2003; 24:77–84. doi: 10.1016/S0197-4580(02)00043-X.
31. Zhang J, Kong Q, Zhang Z, Ge P, Ba D, He W. Telomere dysfunction of lymphocytes in patients with Alzheimer disease. *Cogn Behav Neurol* 2003;16:170–6. doi: 10.1097/00146965-200309000-00004.
32. Pierson E, Simmons SB, Castelli L, Goverman JM. Mechanisms regulating regional localization of inflammation during CNS autoimmunity. *Immunol Rev* 2012; 248:205–15. doi: 10.1111/j.1600-065X.2012.01126.x.
33. Monsonego A, Nemirovsky A, Harpaz I. CD4 T cells in immunity and immunotherapy of Alzheimer's disease. *Immunology* 2013; 139:438–46. doi: 10.1111/imm.12103.
34. Racke MM, Boone LI, Hepburn DL, Parsadainian M, Bryan MT, Ness DK, et al. Exacerbation of cerebral amyloid angiopathy-associated microhemorrhage in amyloid precursor protein transgenic mice by immunotherapy is dependent on antibody recognition of deposited forms of amyloid beta. *JNeurosci* 2005;25:629–36. doi:10.1523/JNEUROSCI.4337-04.2005.
35. Galimberti D, Scarpini E. Inflammation and oxidative damage in Alzheimer's disease: Friend or foe? *Front Biosci (Schol Ed)* 2011; 3:252–66. doi: 10.2741/s149.
36. Sokolova A, Hill MD, Rahimi F, Warden LA, Halliday GM, Shepherd CE. Monocyte chemoattractant protein-1 plays a dominant role in the chronic inflammation observed in Alzheimer's disease. *Brain Pathol* 2009; 19:392–8. doi: 10.1111/j.1750-3639.2008.00188.x.
37. Belkhelda M, Rafa H, Medjeber O, Arroul-Lammali A, Behairi N, Abada-Bendib M, et al. IFN- $\gamma$  and TNF- $\alpha$  are involved during Alzheimer disease progression and correlate with nitric oxide production: A study in Algerian patients. *J Interferon Cytokine Res* 2014; 34:839–47. doi: 10.1089/jir.2013.0085.
38. Lin X, Bai G, Lin L, Wu H, Cai J, Ugen KE, et al. Vaccination induced changes in pro-inflammatory cytokine levels as an early putative biomarker for cognitive improvement in a transgenic mouse model for Alzheimer disease. *Hum Vaccin Immunother* 2014; 10:2024–31. doi: 10.4161/hv.28735.
39. Chakrabarty P, Jansen-West K, Beccard A, Ceballos-Diaz C, Levites Y, Verbeeck C, et al. Massive gliosis induced by interleukin-6 suppresses A $\beta$  deposition in vivo: Evidence against inflammation as a driving force for amyloid deposition. *FASEB J* 2010; 24:548–59. doi: 10.1096/fj.09-141754.
40. Yu JT, Tan L, Song JH, Sun YP, Chen W, Miao D, et al. Interleukin-18 promoter polymorphisms and risk of late onset Alzheimer's disease. *Brain Res* 2009; 1253:169–75. doi: 10.1016/j.brainres.2008.11.083.
41. Felderhoff-Mueser U, Schmidt OI, Oberholzer A, Bührer C, Stahl PE. IL-18: A key player in neuroinflammation and neurodegeneration? *Trends Neurosci* 2005; 28:487–93. doi: 10.1016/j.tins.2005.06.008.
42. Reale M, Kamal MA, Velluto L, Gambi D, Di Nicola M, Greig NH. Relationship between inflammatory mediators, A $\beta$  levels and ApoE genotype in Alzheimer disease. *Curr Alzheimer Res* 2012; 9:447–57. doi: 10.2174/156720512800492549.
43. Conti B, Park LC, Calingasan NY, Kim Y, Kim H, Bae Y, et al. Cultures of astrocytes and microglia express interleukin 18. *Brain Res Mol Brain Res* 1999; 67:46–52. doi: 10.1016/S0169-328X(99)00034-0.
44. Kawanokuchi J, Mizuno T, Takeuchi H, Kato H, Wang J, Mitsuma N, et al. Production of interferon-gamma by microglia. *Mult Scler* 2006; 12:558–64. doi: 10.1177/1352458506070763.
45. Schmitt TL, Steiner E, Klinger P, Sztankay A, Grubeck-Loebenstein B. The production of an amyloidogenic metabolite of the Alzheimer amyloid beta precursor protein (APP) in thyroid cells is stimulated by interleukin 1 beta, but inhibited by interferon gamma. *J Clin Endocrinol Metab* 1996; 81:1666–9. doi: 10.1210/jcem.81.4.8636387.
46. Noguchi A, Nawa M, Aiso S, Okamoto K, Matsuoka M. Transforming growth factor beta2 level is elevated in neurons of Alzheimer's disease brains. *Int J Neurosci* 2010; 120:168–75. doi: 10.3109/00207450903139689.
47. Eslami P, Johnson MF, Terzakaryan E, Chew C, Harris-White ME. TGF beta2-induced changes in LRP-1/T beta R-V and the impact on lysosomal A beta uptake and neurotoxicity. *Brain Res* 2008; 1241:176–87. doi: 10.1016/j.brainres.2008.08.086.
48. Caraci F, Spampinato S, Sortino MA, Bosco P, Battaglia G, Bruno V, et al. Dysfunction of TGF-beta1 signaling in Alzheimer's disease: Perspectives for neuroprotection. *Cell Tissue Res* 2012; 347:291–301. doi: 10.1007/s00441-011-1230-6.
49. Hickman SE, El Khoury J. Mechanisms of mononuclear phagocyte recruitment in Alzheimer's disease. *CNS Neurol Disord Drug Targets* 2010; 9:168–73. doi: 10.2174/187152710791011982.
50. D'Mello C, Le T, Swain MG. Cerebral microglia recruit monocytes into the brain in response to tumor necrosis factor alpha signaling during peripheral organ inflammation. *JNeurosci* 2009; 29:2089–102. doi: 10.1523/JNEUROSCI.3567-08.2009.
51. Selenica ML, Alvarez JA, Nash KR, Lee DC, Cao C, Lin X, et al. Diverse activation of microglia by chemokine (C-C motif) ligand 2 overexpression in brain. *J Neuroinflammation* 2013; 10:86. doi: 10.1186/1742-2094-10-86.
52. Tripathy D, Thirumangalakudi L, Grammas P. RANTES upregulation in the Alzheimer's disease brain: A possible neuroprotective role. *Neurobiol Aging* 2010; 31:8–16. doi: 10.1016/j.neurobiolaging.2008.03.009.
53. Sharma V, Mishra M, Ghosh S, Tewari R, Basu A, Seth P, et al. Modulation of interleukin-1beta mediated inflammatory response in human astrocytes by flavonoids: Implications in neuroprotection. *Brain Res Bull* 2007; 73:55–63. doi: 10.1016/j.brainresbull.2007.01.016.
54. Lumpkins K, Bochicchio GV, Zagol B, Ulloa K, Simard JM, Schaub S, et al. Plasma levels of the beta chemokine regulated upon activation, normal T cell expressed, and secreted (RANTES) correlate with severe brain injury. *J Trauma* 2008; 64:358–61. doi: 10.1097/TA.0b013e318160df9b.
55. Vago L, Nebuloni M, Bonetto S, Pellegrinelli A, Zerbi P, Ferri A, et al. Rantes distribution and cellular localization in the brain of HIV-infected patients. *Clin Neuropathol* 2001; 20:139–45.
56. Laske C, Stellos K, Eschweiler GW, Leyhe T, Gawaz M. Decreased CXCL12 (SDF-1) plasma levels in early Alzheimer's disease: A contribution to a deficient hematopoietic brain support? *J Alzheimers Dis* 2008; 15:83–95.
57. Li M, Hale JS, Rich JN, Ransohoff RM, Lathia JD. Chemokine CXCL12 in neurodegenerative diseases: An SOS signal for stem cell-based repair. *Trends Neurosci* 2012; 35:619–28. doi: 10.1016/j.tins.2012.06.003.

58. Zhu B, Xu D, Deng X, Chen Q, Huang Y, Peng H, et al. CXCL12 enhances human neural progenitor cell survival through a CXCR7- and CXCR4-mediated endocytotic signaling pathway. *Stem Cells* 2012; 30:2571–83. doi: 10.1002/stem.1239.
59. Raman D, Milatovic SZ, Milatovic D, Splittgerber R, Fan GH, Richmond A. Chemokines, macrophage inflammatory protein-2 and stromal cell-derived factor-1 $\alpha$ , suppress amyloid  $\beta$ -induced neurotoxicity. *Toxicol Appl Pharmacol* 2011; 256:300–13. doi: 10.1016/j.taap.2011.06.006.
60. Kim TS, Lim HK, Lee JY, Kim DJ, Park S, Lee C, et al. Changes in the levels of plasma soluble fractalkine in patients with mild cognitive impairment and Alzheimer's disease. *Neurosci Lett* 2008; 436:196–200. doi: 10.1016/j.neulet.2008.03.019.
61. Cho SH, Sun B, Zhou Y, Kauppinen TM, Halabisky B, Wes P, et al. CX3CR1 protein signaling modulates microglial activation and protects against plaque-independent cognitive deficits in a mouse model of Alzheimer disease. *J Biol Chem* 2011; 286:32713–22. doi: 10.1074/jbc.M111.254268.
62. Popp J, Bacher M, Kölsch H, Noelker C, Deuster O, Dodel R, et al. Macrophage migration inhibitory factor in mild cognitive impairment and Alzheimer's disease. *J Psychiatr Res* 2009; 43:749–53. doi: 10.1016/j.jpsychires.2008.10.006.
63. Al-Abed Y, VanPatten S. MIF as a disease target: ISO-1 as a proof-of-concept therapeutic. *Future Med Chem* 2011; 3: 45–63. doi: 10.4155/fmc.10.281.
64. Bacher M, Deuster O, Aljabari B, Egensperger R, Neff F, Jessen F, et al. The role of macrophage migration inhibitory factor in Alzheimer's disease. *Mol Med* 2010; 16:116–21. doi: 10.2119/molmed.2009.00123.
65. Oyama R, Yamamoto H, Titani K. Glutamine synthetase, hemoglobin alpha-chain, and macrophage migration inhibitory factor binding to amyloid beta-protein: Their identification in rat brain by a novel affinity chromatography and in Alzheimer's disease brain by immunoprecipitation. *Biochim Biophys Acta* 2000; 1479:91–102. doi: 10.1016/S0167-4838(00)00057-1.
66. Lashuel HA, Aljabari B, Sigurdsson EM, Metz CN, Leng L, Callaway DJ, et al. Amyloid fibril formation by macrophage migration inhibitory factor. *Biochem Biophys Res Commun* 2005; 338:973–80. doi: 10.1016/j.bbrc.2005.10.040.
67. Xia MQ, Qin SX, Wu LJ, Mackay CR, Hyman BT. Immunohistochemical study of the beta-chemokine receptors CCR3 and CCR5 and their ligands in normal and Alzheimer's disease brains. *Am J Pathol* 1998; 153:31–7.
68. Man SM, Ma YR, Shang DS, Zhao WD, Li B, Guo DW, et al. Peripheral T cells overexpress MIP-1 $\alpha$  to enhance its transendothelial migration in Alzheimer's disease. *Neurobiol Aging* 2007; 28:485–96. doi: 10.1016/j.neurobiolaging.2006.02.013.
69. Li K, Dai D, Yao L, Gu X, Luan K, Tian W, et al. Association between the macrophage inflammatory protein-1 alpha gene polymorphism and Alzheimer's disease in the Chinese population. *Neurosci Lett* 2008; 433:125–8. doi: 10.1016/j.neulet.2008.01.002.
70. Passos GF, Figueiredo CP, Prediger RD, Pandolfo P, Duarte FS, Medeiros R, et al. Role of the macrophage inflammatory protein-1 $\alpha$ /CC chemokine receptor 5 signaling pathway in the neuroinflammatory response and cognitive deficits induced by beta-amyloid peptide. *Am J Pathol* 2009; 175:1586–97. doi: 10.2353/ajpath.2009.081113.
71. Kolev MV, Ruseva MM, Harris CL, Morgan BP, Donev RM. Implication of complement system and its regulators in Alzheimer's disease. *Curr Neuropharmacol* 2009; 7:1–8. doi: 10.2174/157015909787602805.
72. Shen Y, Yang L, Li R. What does complement do in Alzheimer's disease? Old molecules with new insights. *Transl Neurodegener* 2013; 2:21. doi: 10.1186/2047-9158-2-21.
73. Fraser DA, Pisalyaput K, Tenner AJ. C1q enhances microglial clearance of apoptotic neurons and neuronal blebs, and modulates subsequent inflammatory cytokine production. *J Neurochem* 2010; 112:733–43. doi: 10.1111/j.1471-4159.2009.06494.x.
74. Alexander JJ, Anderson AJ, Barnum SR, Stevens B, Tenner AJ. The complement cascade: Yin-Yang in neuroinflammation-neuro-protection and -degeneration. *J Neurochem* 2008; 107:1169–87. doi: 10.1111/j.1471-4159.2008.05668.x.
75. Benoit ME, Hernandez MX, Dinh ML, Benavente F, Vasquez O, Tenner AJ. C1q-induced LRP1B and GPR6 proteins expressed early in Alzheimer disease mouse models, are essential for the C1q-mediated protection against amyloid-beta neurotoxicity. *J Biol Chem* 2013; 288:654–65. doi: 10.1074/jbc.M112.400168.
76. Benoit ME, Tenner AJ. Complement protein C1q-mediated neuroprotection is correlated with regulation of neuronal gene and microRNA expression. *J Neurosci* 2011; 31:3459–69. doi: 10.1523/JNEUROSCI.3932-10.2011.
77. Loeffler DA, Camp DM, Bennett DA. Plaque complement activation and cognitive loss in Alzheimer's disease. *J Neuroinflammation* 2008; 5:9. doi: 10.1186/1742-2094-5-9.
78. Eikelenboom P, Hoozemans JJ, Veerhuis R, van Exel E, Rozemuller AJ, van Gool WA. Whether, when and how chronic inflammation increases the risk of developing late-onset Alzheimer's disease. *Alzheimers Res Ther* 2012; 4:15. doi: 10.1186/alzrt118.
79. Kravitz BA, Corrada MM, Kawas CH. Elevated C-reactive protein levels are associated with prevalent dementia in the oldest-old. *Alzheimers Dement* 2009; 5:318–23. doi: 10.1016/j.jalz.2009.04.1230.
80. Komulainen P, Lakka TA, Kivipelto M, Hassinen M, Penttilä IM, Helkala EL, et al. Serum high sensitivity C-reactive protein and cognitive function in elderly women. *Age Ageing* 2007; 36:443–8. doi: 10.1093/ageing/afm051.
81. Strang F, Scheichl A, Chen YC, Wang X, Htun NM, Bassler N, et al. Amyloid plaques dissociate pentameric to monomeric C-reactive protein: A novel pathomechanism driving cortical inflammation in Alzheimer's disease? *Brain Pathol* 2012; 22:337–46. doi: 10.1111/j.1750-3639.2011.00539.x.
82. Helmy AA, Naseer MM, Shafie SE, Nada MA. Role of interleukin 6 and alpha-globulins in differentiating Alzheimer and vascular dementias. *Neurodegener Dis* 2012; 9:81–6. doi: 10.1159/000329568.
83. O'Bryant SE, Waring SC, Hobson V, Hall JR, Moore CB, Bottiglieri T, et al. Decreased C-reactive protein levels in Alzheimer disease. *J Geriatr Psychiatry Neurol* 2010; 23:49–53. doi: 10.1177/0891988709351832.
84. Sundelöf J, Kilander L, Helmersson J, Larsson A, Rönnemaa E, Degerman-Gunnarsson M, et al. Systemic inflammation and the risk of Alzheimer's disease and dementia: A prospective population-based study. *J Alzheimers Dis* 2009; 18:79–87. doi: 10.3233/JAD-2009-1126.
85. Mohsenzadegan M, Mirshafiey A. The immunopathogenic role of reactive oxygen species in Alzheimer disease. *Iran J Allergy Asthma Immunol* 2012; 11:203–16. doi: 10.1038/ijaai.203216.
86. Shen S, Callaghan D, Juzwik C, Xiong H, Huang P, Zhang W. ABCG2 reduces ROS-mediated toxicity and inflammation: a potential role in Alzheimer's disease. *J Neurochem* 2010; 114:1590–1604. doi: 10.1111/j.1471-4159.2010.06887.x.
87. Xiong H, Callaghan D, Jones A, Bai J, Rasquinha I, Smith C, et al. ABCG2 is upregulated in Alzheimer's brain with cerebral amyloid angiopathy and may act as a gatekeeper at the blood-brain barrier for Abeta(1-40) peptides. *J Neurosci* 2009; 29:5463–75. doi: 10.1523/JNEUROSCI.5103-08.2009.
88. Vladimirova O, Lu FM, Shawver L, Kalman B. The activation of protein kinase C induces higher production of reactive oxygen species by mononuclear cells in patients with multiple sclerosis than in controls. *Inflamm Res* 1999; 48:412–16. doi: 10.1007/s00110050480.

89. Aliev G, Priyadarshini M, Reddy VP, Grieg NH, Kaminsky Y, Cacabelos R, et al. Oxidative stress mediated mitochondrial and vascular lesions as markers in the pathogenesis of Alzheimer disease. *Curr Med Chem* 2014; 21:2208–17. doi: 10.2174/0929867321666131227161303.
90. Cornelius C, Koverech G, Crupi R, Di Paola R, Koverech A, Lodato F, et al. Osteoporosis and Alzheimer pathology: Role of cellular stress response and hormetic redox signaling in aging and bone remodeling. *Front Pharmacol* 2014; 5:120. doi: 10.3389/fphar.2014.00120.
91. Sarti P, Arese M, Giuffre A. The molecular mechanisms by which nitric oxide controls mitochondrial complex IV. *Ital J Biochem* 2003; 52:37–42.
92. Sheng B, Gong K, Niu Y, Liu L, Yan Y, Lu G, et al. Inhibition of gamma-secretase activity reduces Abeta production, reduces oxidative stress, increases mitochondrial activity and leads to reduced vulnerability to apoptosis: Implications for the treatment of Alzheimer's disease. *Free Radic Biol Med* 2009; 46:1362–75. doi: 10.1016/j.freeradbiomed.2009.02.018.
93. Bodovitz S, Klein WL. Cholesterol modulates alpha-secretase cleavage of amyloid precursor protein. *J Biol Chem* 1996; 271:4436–40.
94. Mirshafiey A, Mohsenzadegan M. The role of reactive oxygen species in immunopathogenesis of rheumatoid arthritis. *Iran J Allergy Asthma Immunol* 2008; 7:195–202. doi: 10.1007/ijai.195202.
95. Ferretti MT, Bruno MA, Ducatenzeiler A, Klein WL, Cuervo AC. Intracellular A $\beta$ -oligomers and early inflammation in a model of Alzheimer's disease. *Neurobiol Aging* 2012; 33:1329–42. doi: 10.1016/j.neurobiolaging.2011.01.007.
96. Ferrera P, Arias C. Differential effects of COX inhibitors against beta-amyloid-induced neurotoxicity in human neuroblastoma cells. *Neurochem Int* 2005; 47:589–96. doi: 10.1016/j.neuint.2005.06.012.
97. Lee M, Sparatore A, Del Soldato P, McGeer E, McGeer PL. Hydrogen sulfide-releasing NSAIDs attenuate neuroinflammation induced by microglial and astrocytic activation. *Glia* 2010; 58:103–13. doi: 10.1002/glia.20905.
98. Shi J, Wang Q, Johansson JU, Liang X, Woodling NS, Priyam P, et al. Inflammatory prostaglandin E2 signaling in a mouse model of Alzheimer disease. *Ann Neurol* 2012; 72:788–98. doi: 10.1002/ana.23677.
99. Alzheimer's Association. FDA-Approved treatments for Alzheimer's. From: [www.alz.org/national/documents/topicsheet\\_treatments.pdf](http://www.alz.org/national/documents/topicsheet_treatments.pdf) Accessed: Feb 2015.
100. Zemek F, Drtinova L, Nepovimova E, Sepsova V, Korabecny J, Klimes J, et al. Outcomes of Alzheimer's disease therapy with acetylcholinesterase inhibitors and memantine. *Expert Opin Drug Saf* 2014; 13:759–74. doi: 10.1517/14740338.2014.914168.
101. Wyss-Coray T, Rogers J. Inflammation in Alzheimer disease: A brief review of the basic science and clinical literature. *Cold Spring Harb Perspect Med* 2012; 2:a006346. doi: 10.1101/cshperspect.a006346.
102. McNaull BB, Todd S, McGuinness B, Passmore AP. Inflammation and anti-inflammatory strategies for Alzheimer's disease: A mini-review. *Gerontology* 2010; 56:3–14. doi: 10.1159/000237873.
103. Yu D, Corbett B, Yan Y, Zhang GX, Reinhart P, Cho SJ, et al. Early cerebrovascular inflammation in a transgenic mouse model of Alzheimer's disease. *Neurobiol Aging* 2012; 33:2942–7. doi: 10.1016/j.neurobiolaging.2012.02.023.
104. Leoutsakos JM, Muthen BO, Breitner JC, Lyketsos CG; ADAPT Research Team. Effects of non-steroidal anti-inflammatory drug treatments on cognitive decline vary by phase of pre-clinical Alzheimer disease: Findings from the randomized controlled Alzheimer's Disease Anti-inflammatory Prevention Trial. *Int J Geriatr Psychiatry* 2012; 27:364–74. doi: 10.1002/gps.2723.
105. Jaturapatporn D, Isaac MG, McCleery J, Tabet N. Aspirin, steroidal and non-steroidal anti-inflammatory drugs for the treatment of Alzheimer's disease. *Cochrane Database Syst Rev* 2012; 2:CD006378. doi: 10.1002/14651858.CD006378.pub2.
106. Lemere CA. Immunotherapy for Alzheimer's disease: Hoops and hurdles. *Mol Neurodegener* 2013; 8:36. doi: 10.1186/1750-1326-8-36.
107. Clinical Trials, USA National Institutes of Health. Progress of mild Alzheimer's disease in participants on solanezumab versus placebo (EXPEDITION 3). From: <https://clinicaltrials.gov/ct2/show/NCT01900665> Accessed: Feb 2015.
108. Bateman RJ, Xiong C, Benzinger TL, Fagan AM, Goate A, Fox NC, et al. Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N Engl J Med* 2012; 367:795–804. doi: 10.1056/NEJMoa1202753.
109. Lambracht-Washington D, Rosenberg RN. Advances in the development of vaccines for Alzheimer's disease. *Discov Med* 2013; 15:319–26.
110. Qu B, Boyer PJ, Johnston SA, Hynan LS, Rosenberg RN. Abeta42 gene vaccination reduces brain amyloid plaque burden in transgenic mice. *J Neurol Sci* 2006; 244:151–8.
111. DaSilva KA, Brown ME, McLaurin J. Reduced oligomeric and vascular amyloid-beta following immunization of TgCRND8 mice with an Alzheimer's DNA vaccine. *Vaccine* 2009; 27:1365–76. doi: 10.1016/j.vaccine.2008.12.044.
112. Ubhi K, Masliah E. Alzheimer's disease: Recent advances and future perspectives. *J Alzheimers Dis* 2013; 33:S185–94. doi: 10.3233/JAD-2012-129028.
113. Nixon RA, Wegiel J, Kumar A, Yu WH, Peterhoff C, Cataldo A, et al. Extensive involvement of autophagy in Alzheimer disease: An immuno-electron microscopy study. *J Neuropathol Exp Neurol* 2005; 64:113–22.
114. Yu WH, Cuervo AM, Kumar A, Peterhoff CM, Schmidt SD, Lee JH, et al. Macroautophagy: A novel Beta-amyloid peptide-generating pathway activated in Alzheimer's disease. *J Cell Biol* 2005; 171:87–98. doi: 10.1083/jcb.200505082.
115. Caccamo A, Majumder S, Richardson A, Strong R, Oddo S. Molecular interplay between mammalian target of rapamycin (mTOR), amyloid-beta, and tau: Effects on cognitive impairments. *J Biol Chem* 2010; 285:13107–20. doi: 10.1074/jbc.M110.100420.
116. Martorana A, Balistreri CR, Bulati M, Buffa S, Azzarello DM, Camarda C, et al. Double negative (CD19+IgG+IgD-CD27-) B lymphocytes: A new insight from telomerase in healthy elderly, in centenarian offspring and in Alzheimer's disease patients. *Immunol Lett* 2014; 162:303–9. doi: 10.1016/j.imlet.2014.06.003.
117. Colonna-Romano G, Bulati M, Aquino A, Pellicanò M, Vitello S, Lio D, et al. A double-negative (IgD-CD27-) B cell population is increased in the peripheral blood of elderly people. *Mech Ageing Dev* 2009; 130:681–90. doi: 10.1016/j.mad.2009.08.003.