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## **Inclusion Body Myositis**

### *Navigating diagnostic challenges, case report*

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### **Abstract**

18 Inclusion body myositis (IBM) is a rare progressive myopathy affecting individuals older than 50  
19 years. It is associated with significant morbidity once restricting the patient's mobility, and it has  
20 a relatively low mortality risk with respiratory muscles involvement. Muscle biopsy is the gold  
21 standard method for diagnosis. In this complex scenario, we present a case involving a 72-year-  
22 old woman admitted to our hospital with progressive weakness of lower limbs. The diagnostic  
23 process was challenging due to the case's complexity necessitating a multidisciplinary team  
24 approach. This case highlights the intricate nature of the diagnostic journey, as diagnosing IBM  
25 remains a challenge in clinical practice, requiring a high suspicion and precise application of  
26 available diagnostic tools with the guidance of a collaborative multidisciplinary approach in  
27 investigating and providing patient care. This case report contributes valuable insights to the  
28 understanding of this complex myopathy, facilitating more accurate diagnosis and enhancing  
29 patient care strategies

30 **Keywords:** Sporadic inclusion body myositis; idiopathic inflammatory myopathy; rimmed  
31 vacuoles

32

### 33 **Introduction**

34 Idiopathic inflammatory myopathies are heterogeneous group autoimmune disorders with  
35 varying clinical presentation that affect several systems, including the musculoskeletal,  
36 cardiopulmonary, and gastrointestinal systems, but more prominently skeletal muscles.<sup>1</sup>

37 Inclusion body myositis (IBM) is a subset of the three main idiopathic inflammatory myopathies,  
38 along with others like dermatomyositis and polymyositis.<sup>2,3</sup> It has a slow progression nature with  
39 distinct clinical and pathological presentations. The hallmark of these inflammatory disorders is  
40 inflammation and necrosis of muscle fibers associated with rising levels of muscle enzymes and  
41 presenting primarily as weakness. IBM is of two types, sporadic and hereditary; both have  
42 similar features with one distinction: the absence of inflammation in the latter.<sup>4,5</sup>

43

44 The prevalence of IBM is frequently underestimated due to diagnostic challenges and a high rate  
45 of misdiagnosis. A study in the Netherlands estimated a prevalence of 5 cases per million in  
46 2000.<sup>6</sup> But more recently, a population-based study in Ireland in 2017 reported a much higher  
47 prevalence of 112 cases per million, and a recent meta-analysis reported a pooled prevalence of  
48 46 patients per million, reflecting a sharp increase in the last decade, indicating a significant  
49 increase possibly attributed to improved diagnostic methods and increased awareness.<sup>6,8</sup>

50

51 The importance of accurate classification became apparent with the incorporation of IBM into  
52 the International Classification of Diseases (ICD). In 2018, the introduction of the ICD, Ninth  
53 Revision, and Clinical Modification (ICD9CM) code offered a more precise estimation of  
54 prevalence and health care costs, revealing an annual cost of \$35,000 for patients with IBM and  
55 Medicare coverage, alongside a prevalence of 84 cases per million in individuals over 65 years  
56 of age in the United States.<sup>7,9</sup>

57

58 Inclusion body myositis is more common in males than in females and most patients progress to  
59 being wheelchair bound by 20 years from the first presentation.<sup>5</sup> Classically, there is an  
60 asymmetric involvement of finger flexors and knee extensors. However, atypically, dysphagia,

61 weakness in the proximal upper limbs, or axial muscles may also occur, particularly in advanced  
62 cases. The eventual involvement of respiratory muscles is anticipated, contributing to premature  
63 mortality.<sup>5</sup> Bio-chemically, abnormal creatinine kinase levels and the detection of monoclonal  
64 immunoglobulin through serum immunofixation may be observed. Additionally, positive results  
65 may be noted for other markers, including antinuclear antibodies, anti-RO antibodies, anti-La  
66 antibodies, rheumatoid factor, and anti-cN1A autoantibody.<sup>7</sup> The presentation of our patient  
67 mirrored the classical features of IBM. However, the presence of concurrent medical issues  
68 posed a diagnostic challenge, necessitating the application of various diagnostic tools to arrive to  
69 the final diagnosis.

70

### 71 **Case Report**

72 A 72-year-old female patient, with a medical history that includes type 2 diabetes mellitus,  
73 hypertension, and dyslipidaemia, is currently receiving treatment with metformin for her diabetes  
74 and valsartan for her hypertension. She presented with a 3-month history of progressive lower  
75 limb weakness, inability to stand or walk with gradual loss of mobility. Her symptoms were  
76 progressive, eventually rendering her bedbound with a decline in performing daily activities.  
77 Additionally, occasional episodes of disorientation and visual hallucinations were reported.  
78 There was no history of fever, convulsions, upper respiratory symptoms, dysuria, or any rashes.  
79 There was no significant family history of autoimmune disorders or malignancies and no  
80 contributing environmental exposures.

81

82 On initial examination, the patient was alert, communicative, and following simple commands.  
83 While no fasciculations were observed, there was mild muscle wasting in the lower limbs with  
84 evident hypotonia. Assessment of muscle power revealed the following: proximal upper limb  
85 power ranged from 1-2/5, elbow flexion at 2-3/5, and wrist and hand grip at 3/5. Proximal  
86 muscles in the lower limbs exhibited power of 1/5, with knee flexion at 1/5 and  
87 dorsi/plantarflexion of 3/5. Deep tendon reflexes were diminished, and the plantar response was  
88 normal, with no evidence of sensory loss.

89

90 Her Initial laboratory workup (Table 1) showed normal inflammatory markers but exhibited  
91 elevated liver enzymes. Renal parameters and electrolytes were within normal apart from

92 hyponatremia. Her creatine kinase levels were normal on two separate occasions. The  
93 autoimmune myositis screening returned a strong positive result for anti-Ku antibodies. Other  
94 autoimmune parameters, including antinuclear antibodies (ANA) and antineutrophil cytoplasmic  
95 antibodies (ANCA), were negative. She also had a positive latent syphilis profile. Lumbar  
96 puncture results including limbic encephalitis screening and screening for paraneoplastic  
97 syndrome were unremarkable.

98  
99 Neuroimaging, namely computed tomography, and magnetic resonance imaging of the brain  
100 showed features consistent with cerebral atrophy and microvacuolar changes.

101  
102 Patient nerve conduction study showed mild prolongation of distal latencies, slowing of the  
103 conduction velocities, and preserved compound muscle action potential amplitude of bilateral  
104 ulnar, peroneal, and tibial motor neurons. Her electromyography (EMG) showed active  
105 denervation of the examined muscles with mixed myopathic and neurogenic units and reduced  
106 recruitment. Her electroencephalogram tracings showed global slowing with occasional  
107 generalized bursts of high amplitude slow waves consistent with cortical dysfunction.

108  
109 After performing EMG and the nerve conduction study, a muscle biopsy of the left quadriceps  
110 femoris was performed, based on the EMG report. Her muscle biopsy showed numerous  
111 regenerating myofibers, dispersed necrotic/degenerate myofibers, and significant numbers of  
112 myofibers with striking rimmed vacuoles (Figure 1.). There was Type II myofiber atrophy and  
113 increased variation in the size of Type I myofibers. Foci of mild endomysial and perivascular  
114 lymphocytic inflammation were noted and there was mild increased perimysial fat infiltration.  
115 Immunohistochemistry showed sarcolemma upregulation of MHC-1 stain. MAC stain (C5b-9)  
116 was positive in necrotic myofibers but negative in capillaries. The inflammatory cells consisted  
117 mainly of many CD68+ macrophages and occasional endomysial CD3+ T-lymphocytes  
118 comprised of CD4+ > CD8+ lymphocytes (the latter also noted in necrotic fibers). There was  
119 also strong p62 immunoreactivity in scattered rimmed vacuoles (Figure 2.) and in small protein  
120 aggregates within myofibers.

121

122 As part of work up, 18-Fluoro-deoxyglucose positron emission tomography (FDG-PET),  
123 (FIGURE 3.) was performed. The images showed a low-grade homogenous increased FDG  
124 uptake involving the muscles and subcutaneous tissue of the left lateral thigh, worse on the  
125 vastus lateralis, these findings are consistent with myositis.

126

127 Considering the initial suspicion of inflammatory myopathy, the patient was commenced on  
128 treatment courses of intravenous methylprednisolone and intravenous immunoglobulins,  
129 followed by Rituximab injections and oral prednisolone. However, in view of the absence of  
130 autoimmune symptoms, the negative results for ANA and ANCA, along with the possibility of  
131 false-positive anti-Ku antibodies and the diagnostic findings from the muscle biopsy, a diagnosis  
132 IBM was established. Additionally, and following the confirmation of the diagnosis, the patient's  
133 treatment was overseen by a collaborative, multidisciplinary team, from general medicine,  
134 neurology, infectious diseases, physiotherapy, and speech therapy. The treatment plan involved  
135 the initiation of Tenofovir for chronic Hepatitis B and benzylpenicillin for syphilis in view of  
136 needing to continue immunosuppressant medications. Continuous physiotherapy was strongly  
137 emphasized.

138

139 During her hospital stay, the patient showed a moderate clinical improvement, characterized by  
140 enhanced trunk stability and the capacity to maintain evaluated postures. Additionally, there was  
141 a significant increase in muscular strength across all limbs. Subsequently, she was discharged to  
142 her home, with arrangements made for continued treatment at a tertiary care hospital. A verbal  
143 consent was taken from patient's next of kin to report the case.

144

## 145 **Discussion**

146 This case represents exceptional challenges due to the patient's complex medical history,  
147 including hyponatremia, syndrome of inappropriate antidiuretic hormone, a positive syphilis  
148 profile, hypercalcemia, hepatic encephalopathy, and initial suspicion of paraneoplastic  
149 syndrome. Investigating these concurrent conditions significantly prolonged the diagnostic  
150 process, as her symptoms could easily have been misconstrued as manifestations of any of these  
151 underlying issues and they can influence the disease progression. Our report explores the

152 challenges in diagnosing and managing IBM requiring a collaborative effort of a  
153 multidisciplinary team approach for diagnosis and management.

154  
155 Pathogenesis of IBM has been debated due to its unclear nature. Published literature has shown  
156 that IBM results from a series of immune and degenerative reactions with no specific trigger.  
157 Auto-immunity is believed to have a significant role; however, inflammation is the primary  
158 precipitating factor of this cascade.<sup>10</sup> The pathogenesis is thought to revolve around multiple  
159 processes namely rimmed vacuoles and myonuclear degeneration, mitochondrial pathology, and  
160 protein aggregation. One important concept is the immune process led by the infiltration of  
161 CD+8 T cells within muscles triggered by inflammation, production of differentiated cells results  
162 which further exerts its cytotoxic properties causing the production of autoantibodies like cN1A  
163 which is detected in 60% of patients. Moreover, another nonimmune process that works in  
164 parallel is the production of gamma interferon which results in the accumulation of protein  
165 aggregates.<sup>10</sup> Several distinctive molecules were described previously in the pathogenesis of  
166 IBM like amyloid seen by congo red, ubiquitin, B-amyloid, and tau. But more recently,  
167 degenerative muscle biomarkers like p62, LC4, and TDP43 proved to be more valuable and  
168 superior in the detection of IBM.<sup>7</sup> Lastly, mitochondrial dysfunction due to inflammatory  
169 cytokines also results in mitochondrial damage through oxidative stress, the severity of this  
170 damage is correlated to atrophy of muscle fibers.<sup>4,10</sup> Genetic predisposition with the presence of  
171 some HLA genes is presumed to have a part in sporadic IBM.<sup>4</sup>

172  
173 Diagnosis of IBM is highly dependent on histopathological findings which are reflective of the  
174 disease process. Commonly presence of rimmed vacuoles, protein deposits, CD8+ T cells, and  
175 major histocompatibility complex are diagnostic. The latter could also present in polymyositis,  
176 dermatomyositis, and muscular dystrophies.<sup>10</sup> The decision of which muscle to biopsy should be  
177 taken cautiously to avoid false negative results.<sup>11</sup>

178  
179 Electromyography is done as part of investigations; the results can be used to select the site of  
180 biopsy. Both long-duration high amplitude and short-duration and low amplitudes might be  
181 present imposing a challenge in result analysis.<sup>11</sup>

182

183 Musculoskeletal radiology is now emerging as a non-invasive modality deemed useful in  
184 detecting specific patterns associated with IBM distinguishing it over the remaining myositis.  
185 For instance, Magnetic resonance might demonstrate features like muscle atrophy, edema, and  
186 fatty infiltration which assist in biopsy site selection, monitoring disease progression and  
187 differentiating IBM from the other subsets of myositis. In a recent review, the distinct features  
188 seen in different imaging modalities were summarized to help in diagnosing IBM. For example,  
189 in PET scans using F-18 FDG, specific tracers are used to detect potential markers like Beta  
190 amyloid and tau proteins in affected muscles and to monitor the progression of the disease.  
191 Moreover, dual energy x-ray absorptiometry (DEXA) scan can demonstrate muscle mass and  
192 quantities of muscle atrophy, which ultimately helps assess disease progression or evaluate  
193 therapeutic effects.<sup>12</sup>

194

195 Non-Pharmacological management of inclusion body myositis includes measures towards  
196 ensuring regular assessment of swallowing and respiratory muscles along with tackling mobility  
197 and ambulation through exercise routines. Fall precautions and education should also be  
198 emphasized.<sup>7,11</sup>

199

200 Swallowing dysfunction, falls, and a decline in quality of life are common in IBM and  
201 significantly affect morbidity and mortality. Around 40% of patients complain of dysphagia at  
202 diagnosis and around 80% have dysphagia with the advancement of disease. For evaluation of  
203 dysphagia in IBM, rosenbek penetration aspiration scale, video-fluoroscopy, and endoscopic  
204 evaluation of swallowing along with magnetic resonance can quantify swallowing dysfunction.  
205 Several measures like the Mendelsohn maneuver and expiratory muscle strength trainer device to  
206 reduce the risk of aspiration were proposed for prevention, however, no role was identified for  
207 improving swallowing function. Similarly, ankle foot orthosis has been proposed for prevention  
208 of recurrent falls which correlate with disease progression, however a definitive role in the  
209 prevention of falls has not been established.<sup>13</sup>

210

211 The role of immunosuppression therapy has transient effects with myodegeneration being the  
212 main target for therapy, especially in cases where bulbar and proximal muscle presentation are

213 exhibited. Agents like arimoclomol, bimagrumab, follistatin, oxandrolone, and rapamycin were  
214 proposed in recent clinical trials.<sup>3</sup>

215  
216 In one study (n=43), follow up of patients over a period averaging 61.1 months revealed a  
217 monthly muscle power deterioration of -0.79%. The natural course without immunosuppressive  
218 treatment showed a steeper decline at -1.03% per month most rapidly at knee extension. Males in  
219 the initial 5 years post-onset saw a quicker decline, notably, serum creatine kinase levels, region  
220 and age at onset didn't predict prognosis. Inversely, treated patients had a significantly lower  
221 decline (-0.76%) than untreated (-1.03%), and mycophenolate mofetil treatment showed a more  
222 favorable prognosis at -0.67%. These findings emphasize the potential benefit of  
223 immunosuppressive interventions in slowing muscle power loss in IBM patients.<sup>14</sup>

224

## 225 **Conclusion**

226 The diagnosis of IBM necessitates a comprehensive approach that integrates clinical evaluation,  
227 biochemical analysis, and metabolic imaging such as F-18 FDG PET-CT. In this case, we not  
228 only emphasized the significance of this multimodality approach and clinical acumen but also  
229 emphasize the vital role of multidisciplinary management. The collaborative efforts of experts  
230 from diverse fields, including general medicine, neurology, pathology, radiology, and  
231 rehabilitation therapy, were instrumental in not only arriving at an accurate diagnosis but also  
232 formulating a holistic management plan tailored to the patient's specific need.

233

## 234 **Authors' Contribution**

235 Drafting the manuscript was done by MS and SB. SJ provided the details of histopathology  
236 slides and description. JZ provided details of imaging and related description. HF managed the  
237 case. AA managed the case and critically revised the manuscript. All authors approved the final  
238 version of the manuscript.

239

## 240 **References**

- 241 1. Lundberg IE, Fujimoto M, Vencovsky J, Aggarwal R, Holmqvist M, Christopher-Stine L,  
242 et al. Idiopathic inflammatory myopathies. *Nat Rev Dis Primers*. 2021;7(1):86.  
243 <https://doi.org/10.1038/s41572-021-00321-x>



- 244 2. Oldroyd A, Lilleker J, Chinoy H. Idiopathic inflammatory myopathies - a guide to  
245 subtypes, diagnostic approach and treatment. *Clin Med (Lond)*. 2017;17(4):322-8.  
246 <https://doi.org/10.7861/clinmedicine.17-4-322>
- 247 3. Connolly CM, Plomp L, Paik JJ, Allenbach Y. Possible future avenues for myositis  
248 therapeutics: DM, IMNM and IBM. *Best Pract Res Clin Rheumatol*. 2022;36(2):101762.  
249 <https://doi.org/10.1016/j.berh.2022.101762>
- 250 4. Catalan M, Selva-O'Callaghan A, Grau JM. Diagnosis and classification of sporadic  
251 inclusion body myositis (sIBM). *Autoimmun Rev*. 2014;13(4-5):363-6.  
252 <https://doi.org/10.1016/j.autrev.2014.01.016>
- 253 5. Naddaf E. Inclusion body myositis: Update on the diagnostic and therapeutic landscape.  
254 *Front Neurol*. 2022;13:1020113. <https://doi.org/10.3389/fneur.2022.1020113>
- 255 6. Badrising UA, Maat-Schieman M, van Duinen SG, Breedveld F, van Doorn P, van  
256 Engelen B, et al. Epidemiology of inclusion body myositis in the Netherlands: a nationwide  
257 study. *Neurology*. 2000;55(9):1385-7. <https://doi.org/10.1212/wnl.55.9.1385>
- 258 7. Greenberg SA. Inclusion body myositis: clinical features and pathogenesis. *Nat Rev*  
259 *Rheumatol*. 2019;15(5):257-72. <https://doi.org/10.1038/s41584-019-0186-x>
- 260 8. Lefter S, Hardiman O, Ryan AM. A population-based epidemiologic study of adult  
261 neuromuscular disease in the Republic of Ireland. *Neurology*. 2017;88(3):304-13.  
262 <https://doi.org/10.1212/wnl.0000000000003504>
- 263 9. Keshishian A, Greenberg SA, Agashivala N, Baser O, Johnson K. Health care costs and  
264 comorbidities for patients with inclusion body myositis. *Curr Med Res Opin*. 2018;34(9):1679-  
265 85. <https://doi.org/10.1080/03007995.2018.1486294>
- 266 10. Nagy S, Khan A, Machado PM, Houlden H. Inclusion body myositis: from genetics to  
267 clinical trials. *J Neurol*. 2023;270(3):1787-97. <https://doi.org/10.1007/s00415-022-11459-3>
- 268 11. Skolka MP, Naddaf E. Exploring challenges in the management and treatment of  
269 inclusion body myositis. *Curr Opin Rheumatol*. 2023;35(6):404-13.  
270 <https://doi.org/10.1097/bor.0000000000000958>
- 271 12. Goyal NA, Mozaffar T, Dimachkie MM. Imaging beyond muscle magnetic resonance  
272 imaging in inclusion body myositis. *Clin Exp Rheumatol*. 2023;41(2):386-92.  
273 <https://doi.org/10.55563/clinexprheumatol/uimkey>

- 274 13. Ma AK, Dai F, Roy B. In-patient comorbidities in inclusion body myositis: a United  
 275 States national in-patient sample-based study. Clin Exp Rheumatol. 2023;41(2):261-6.  
 276 <https://doi.org/10.55563/clinexprheumatol/791fq8>
- 277 14. Lindberg C, Oldfors A. Prognosis and prognostic factors in sporadic inclusion body  
 278 myositis. Acta Neurol Scand. 2012;125(5):353-8. [https://doi.org/10.1111/j.1600-](https://doi.org/10.1111/j.1600-0404.2011.01584.x)  
 279 0404.2011.01584.x

280  
 281 **Table 1:** Laboratory results values with normal ranges

Test	Result Value	Normal ranges
Total white cell count	10.7 10 <sup>9</sup> /L	2.4-9.5 10 <sup>9</sup> /L
Neutrophils	10 10 <sup>9</sup> /L	1-4.8 10 <sup>9</sup> /L
Lymphocytes	0.4 10 <sup>9</sup> /L	1.2-3.8 10 <sup>9</sup> /L
Hemoglobin	10 g/dL	11-14.5. g/dL
Serum Sodium	126 mmol/L	135-145mmol/L
Serum Calcium	2.68 mmol/L	1.15-2.55 mmol/L
Creatine kinase	54 U/L	26-192 U/L
Osmolality in serum	248 mOsm/kg	275-295 mOsmol/kg
Fractional excretion of sodium	124 mmol/L	135-145 mmol/L
Osmolality in urine	570 mOsm/kg	40-1400 mOsmol/kg
Cortisol	298 nmol/L	am 133-537 nmol/L
Alanine aminotransferase	Day of admission: 35 Day 10 of admission 136 Last reading before discharge 46	0-33 U/L
Aspartate aminotransferase	Day of admission: 20 Day 10 of admission 62 Last reading before discharge 28	0-32 U/L
Alkaline phosphatase	Day of admission: 103 Day 10 of admission:125 Last reading before discharge 97	35-104 U/L
Bilirubin	Day of admission: 9 Day 10 of admission:10 Last reading before discharge :5	0-17umol/L
Limbic encephalitis screen in Cerebrospinal fluid	Negative	-
Paraneoplastic syndrome screen in Cerebrospinal fluid and serum	Negative	-

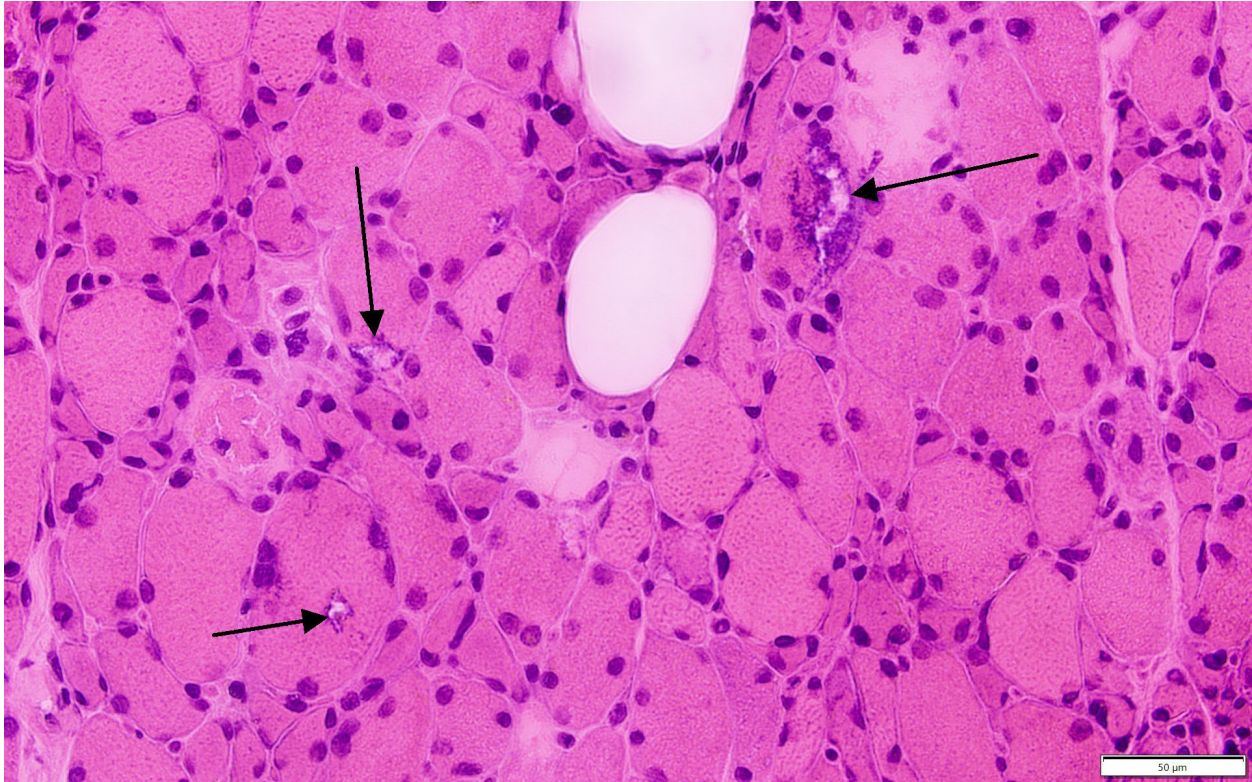
Autoimmune myositis screen	Strong positive Anti-Ku remaining antibodies are negative.	List of tested antibodies: Anti-Mi-2 alpha, Anti-Mi-2 beta, Anti-TIF1f, Anti-MDA5, Anti-NXP2, Anti-SAE1, Anti-Ku, Anti-PM-Scl100, Anti-PM-Scl175,, Anti-Jo-1, Anti-SRP, Anti-PL-7, Anti-PL-12, Anti-EJ, Anti-OJ Anti-Ro52
Anti-nuclear antibody	Negative	
Anti-neutrophilic cytoplasmic antibody	Weak positive c-ANCA - anti-myeloperoxidase MPO: 1 U/ml -anti-proteinase 3 antibodies PR3: 1 U/ml	0.00-20.00 U/ml
Thyroid antibody	136 IU/ml	0-50 IU/ml
Free thyroxin T4	20.5 pmol/L	12.3-20.2 pmol/L
Thyroid stimulating hormone	4.59 mIU/L	0.27-4.20 mIU/L
Parathyroid hormone	6.5 pmol/L	1.6-6.9 pmol/L
Syphilis	Negative rapid plasma regain, non-reactive venereal disease research laboratory. Positive treponema pallidum hemagglutination with a titer of 320	
Anti-Hepatitis B core	Positive	
HCV antibodies	Negative	
Protein electrophoresis, serum	Immunoglobulin G 41.9 g/L remaining immunoglobulin within normal. No abnormal protein bands were detected and confirmed by IFE	7-16g/L
Protein electrophoresis, urine	Urine protein 0.70 g/L	0.00-0.15 g/L
Free light chain profile	Normal	

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287 **Figure 1:** H&E stain of frozen skeletal muscle tissue showing variation in myofiber size and  
288 striking rimmed vacuoles in occasional myofibers (black arrows) [magnification= x400].

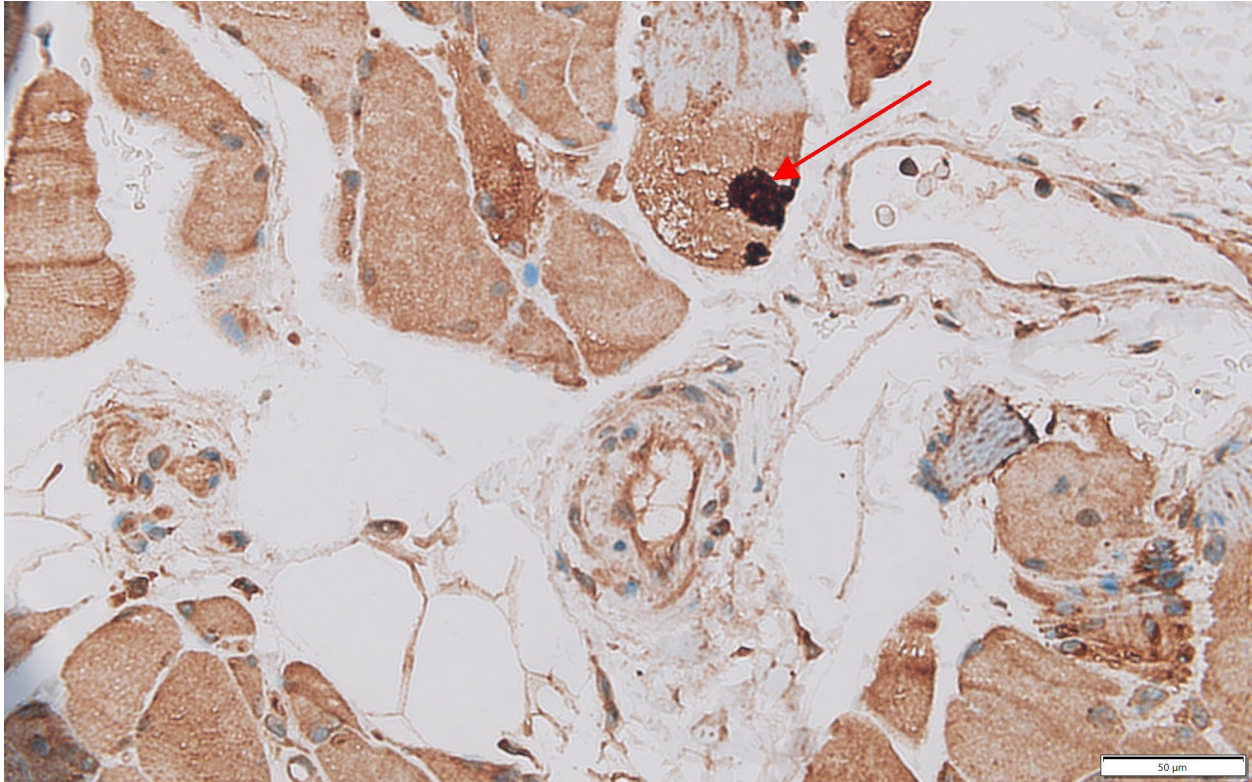
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295 **Figure 2:** Immunohistochemical stain of skeletal muscle tissue showing strong positive  
296 immunoreactivity of **p62** in a rimmed vacuole (red arrow) [magnification= x600].

297

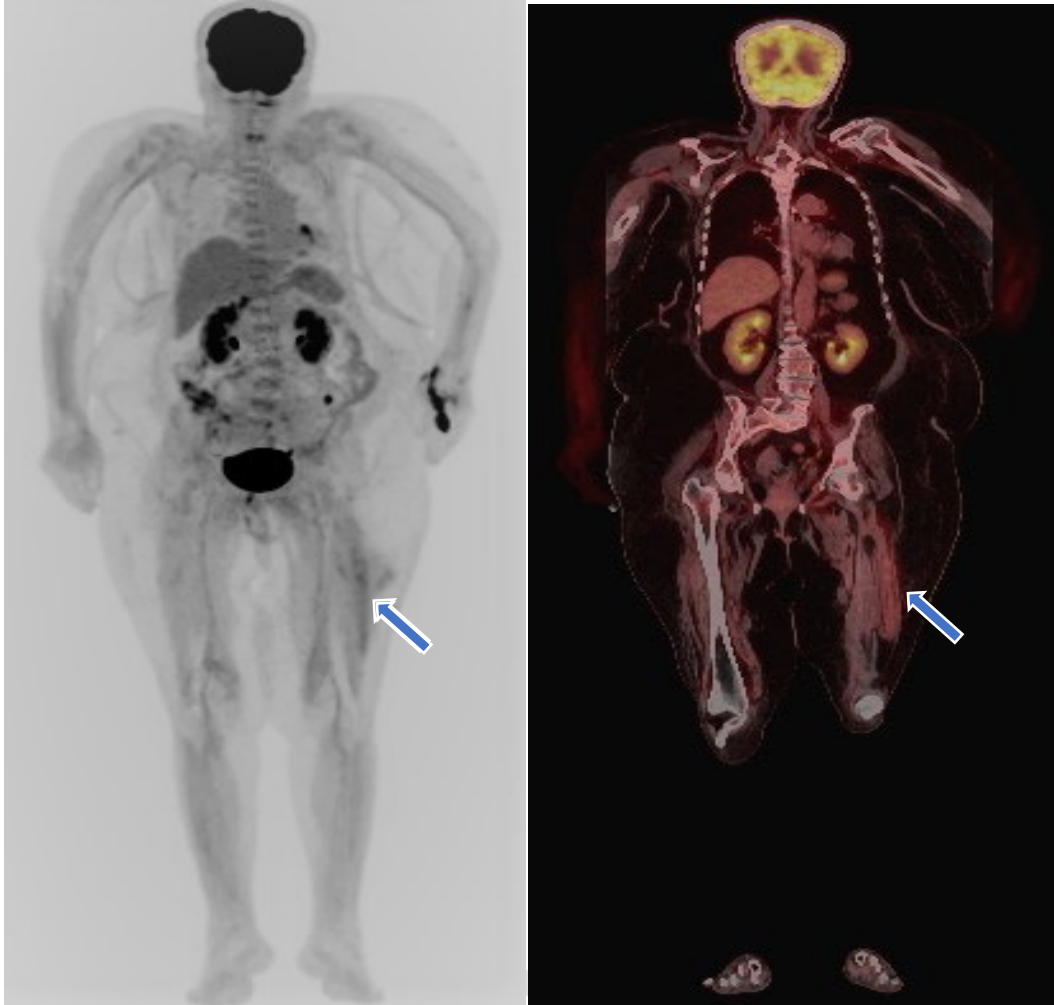
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304 **Figure 3:** F-18 FDG PET-CT: showing low grade homogenous increased FDG uptake involving  
305 the muscles and subcutaneous tissue of the left lateral thigh, worse on the vastus lateralis, these  
306 findings are consistent with myositis (see arrow).

307

308